

SUPPLEMENTARY MATERIAL (updated 12-8-2019) of:

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Literature on free radical generation after exposure to static and extremely-low-frequency electromagnetic fields (as of July 2019)

Two types of free radicals can be generated: reactive oxygen species (ROS) and reactive nitrogen species (RNS). Activity in the mitochondrial electron transport chain leads to the production of superoxide radical anion ($O_2^{\cdot-}$) which can be converted to hydrogen peroxide (H_2O_2) by various forms of superoxide dismutase (SOD). H_2O_2 can be degraded by catalase (CAT) into water and oxygen or converted by the iron-dependent Fenton reaction into the potent hydroxyl radical (OH^{\cdot}). In the cytoplasm, nitric oxide (NO^{\cdot}) is generated by various forms of nitric oxide synthase (NOS) by conversion of L-arginine to L-citrulline. NO^{\cdot} reacts with $O_2^{\cdot-}$ to generate the potent oxidant peroxynitrite ($ONOO^{\cdot}$). $O_2^{\cdot-}$ can also be produced by NOS by transfer of electron from NADPH to O_2 . Other enzymatic processes, such as cytochrome P_{450} , also generate ROS in normal cellular activities.

Major anti-oxidative processes in cells include catalase/peroxidase that converts $O_2^{\cdot-}$ to H_2O and O_2 . In the process, glutathione (GSH) is oxidized to glutathione disulfide (GSSG). GSSG is reduced back to GSH by the enzyme glutathione reductase with the conversion of NADPH to NADP. GSH and NADPH are the most common electron donors participated in cellular anti-oxidation processes. $ONOO^{\cdot}$ is decomposed by peroxiredoxin and glutathione peroxidase into less potent nitrogen free radicals ($NO_3^{\cdot}/NO_2^{\cdot}$).

ROS react with cellular macromolecules, e.g., DNA, protein, and lipid. The most common form of DNA oxidative damage is the formation of hydroxylated bases. 8-hydroxy-2'-deoxyguanosine (8-OHdG) is generally used as an index of oxidative DNA damage. ROS react with lipids to produce lipid peroxy radicals and lipid hydroperoxides. Lipid peroxy can subsequently form malondialdehyde (MDA), which is commonly used as an index of oxidative lipid damage. Lipid radicals can diffuse through membrane leading to protein oxidation and formation of DNA-MDA adduct. Oxidative lipid damages affect the structure and function of cell membrane. ROS attack proteins directly and indirectly. Protein carbonyl is a form of protein oxidative damage. Changes in protein structure lead to alteration in enzymatic activities, particularly, damage to membrane transport proteins leads to ionic imbalance such as intracellular concentrations of calcium and potassium. Oxidative stress could also cause changes in regulation of transcription factors in cells, e.g., the Nrf2 antioxidant pathway.

More than 200 papers have been published on effects of *in vitro* and *in vivo* exposure to static and extremely–low frequency electromagnetic fields on various aspects of the free radical processes in living organisms. Table 1 is a summary of these papers (as of April 2019). (It is inevitable that some relevant studies were omitted in the following literature survey.)

Table 1. Summary of papers on the effects of ELF-EMF on oxidative processes in cells and animals. (* Study reported no significant effect on oxidative processes; □ increase; ▢ decrease; ∅ no significant effect; MF= magnetic field; EF = electric field; CAT= catalase; GSH= glutathione; GST = glutathione S-transferase; GPx = glutathione peroxidase; NOS= nitric oxide synthase; MPO= myeloperoxidase; ROS = reactive oxygen species; SOD= superoxide dismutase) In some studies, the term EMF (electromagnetic field) was used. The authors may mean magnetic field or a combination of magnetic and electric fields, since most exposure systems emit both fields when not properly shielded and grounded. On the other hand, fields labelled as magnetic field in some studies may contain also electric component .,

			<i>Oxidative damages (DNA, protein, lipid)</i>	<i>ROS (O₂⁻, OH, H₂O₂, NO)</i>	<i>NOS</i>	<i>Antioxidative processes (SOD, CAT/ peroxidase, GSH, GPx)</i>	<i>Effect of antioxidants / scavengers</i>	<i>Remarks</i>
Akan et al. (2010)	Activated THP-1 cells (human monocytic leukemia cells)	50-Hz EMF, 1 mT, 4-6 h		□NO	□iNOS			□cGMP
Akdag et al. (2007)	Sprague-Dawley rat serum in vivo	50-Hz MF, 0.1 and 0.5 mT, 2 h/day, 10 months		□NO				
Akdag et al. (2010)	Sprague-Dawley rat brain in vivo	50-Hz MF, 0.1 and 0.5 mT, 2 h/day, 10 months	□ lipid peroxidation			□ CAT		□ total oxidant status, □ total anti-oxidative capacity
Akdag et al. (2013a)	Sprague-Dawley rat brain in vivo	50-Hz MF, 0.1 and 0.5 mT, 2 h/day, 10 months	□ protein carboxylation □ lipid peroxidation					
*Akdag et al. (2013b)	Sprague-Dawley rat testes in vivo	50-Hz MF, 0.1 and 0.5 mT, 2 h/day, 10 months	∅ lipid peroxidation			∅ CAT		No change in total oxidant status and total anti-oxidative capacity
Akpınar et al.	Wister rat brain and	50-Hz EF, 12 and 18	□ lipid peroxidation					□ total oxidant

(2012)	retina in vivo	kV/ m, 1 h/day, 14 days						status, □ total anti-oxidative capacity
Akpinar et al. (2016)	Wister rat brain in vivo	50-Hz EF, 12kV/ m, 1 h/day, prenatal (Pr), postnatal (Po, 30 days), and prenatal + postnatal (PP)	□lipid peroxidation in Po, □in PP (cf. Pr and Po) □protein carboxylation in PP					
Aksen et al. (2006)	Wister rat uterus and ovary in vivo	50-Hz EMF, 1 mT, 3 h/day, 50 or 100 days	□ lipid peroxidation					
*Alcaraz et al. (2014)	Micronucleated cells induced by EMF in bone marrow of mouse	50-Hz EMF, 0.2 mT, for 7, 14, 21, or 28 days					Effect not blocked by 4 types of antioxidant	
Amara et al. (2009)	Male Wistar rat, frontal cortex and hippocampus	Static MF, 128 mT, 1 h/day, 30 days	∅ DNA			□ GPx, CuZn-SOD and CAT in frontal cortex ; □ CuZn-SOD and Mn-SOD in hippocampus		
Amara et al. (2011)	Male Wistar rat, frontal cortex	Static MF, 128 mT, 1 h/day, 30 days and cadmium treatment	Synergistic with cadmium in □lipid peroxidation			Synergistic with cadmium in □SOD and glutathione		
Ansari et al. (2016)	NMRI mice	50-Hz MF, 0.5 mT, 2 h or 2 h/day for 2 weeks		□NO				Blocked effect of L-NAME, a NOS inhibitor, long ter exposure reversed depressive disorder in mice
Asghar et al. (2016)	Soybean seeds and seedlings	50-Hz MF, 50, 75, or 100 mT for	□ lipid peroxidation at 50 mT for 3 min (not at other exposure	□H ₂ O ₂ at 50 and 100 mT for 3 min		□ SOD at 75 mT for 3 and 5 min; □CAT/ peroxidase at 50, 75 and 100 mT for 3 min;		□ ascorbic acid

		3 or 5 min	conditions)					
Ayşe et al. (2010)	K562 cells, in vitro	50-Hz EMF, 5 mT, 1 h or 1 h/day for 4 days		\square O ₂ ^{•-}				Effect disappeared at 2 h post-exposure, no interaction with hemin
Bawin et al. (1996)	Electrical activity of rat hippocampal slices	1-Hz MF, 0.56 and 0.056 mT, 10 min			Effect blocked by NOS inhibitor			60-Hz MF has no significant effect
Bediz et al. (2006)	Sprague-Dawley rat blood and brain in vivo	50-Hz EMF, 0.005 mT, 5 min every other day for 6 months	\square lipid peroxidation			\square GSH		Effect attenuated by zinc
Belova et al. (2010)	Activated mouse peritoneal neutrophils	Combined magnetic field (CMF) tuned to calcium ion (DC 40.6 μ T, AC 74.7 μ T at 31 Hz): pulsed MF (225 μ s, 20 pulses packet at 15 Hz, 1500 μ T); up to 30 min exposure		CMF \square ROS, pulsed MF \square ROS				
Benassi et al. (2016)	SH-SY5Y cells (human used to study Parkinson's disease)	50-Hz MF, 1 mT, 6-72 h	\square protein carboxylation					
Bertea et al. (2015)	Arabidopsis thaliana	Reversed geomagnetic field, 10 days during seed growth						Retarded root and leaf growth, changes in expression of 5 gene

								involved in oxidative stress
Buczyński et al. (2005)	Human blood platelets	1 kHz MF, 0.5 mT, 30, 60 or 90 min	□ lipid peroxidation					Effect observed only after 30 and 90 min exposure, not at 60 min
Budziosz et al. (2018)	Male Wistar rats, frontal cortex, hippocampus, brainstem, hypothalamus, striatum, cerebellum	50-Hz EMF, 22 h/day, 28 days, 4.4 pT	∅ lipid peroxidation	∅ total oxidant status		Changes (□ and □) in SOD, CAT and glutathione-related enzymes depended on brain region		
Buldak et al. (2012)	AT478 murine squamous carcinoma cells	EMF 50-Hz, 1 mT, 16 min	□ lipid peroxidation			□SOD □GPx		MF lessens oxidative effects of cisplatin
Calabro et al. (2013)	SH-SY5Y cells	Static MF, 2.2 mT, 24 h		□ ROS production				
Calcabrini et al. (2017)	Human keratinocyte (NCTC 2544)	50 Hz MF, 0.025 – 0.2 mT, 1 h	□ lipid peroxidation at 0.05 and 0.1 mT	□ ROS at 0.05 and 0.1 mT		□ SOD and □GSH at 0.05 and 0.1 mT		□ ROS blocked by the iron chelator o-phenanthroline
Calota et al. (2006)	Human blood serum	50-Hz EF, 5, 7.5 10, 15, 20 kV/m, 1-2 h		□ ROS production				
Calota et al. (2007)	Human blood serum	50-Hz MF, 0.357, 0.596, 1.788, 2.384 mT, 1-2 h		□ ROS production, enhanced by FeCl ₂ and H ₂ O ₂				
Canseve	Guinea pig,	50-Hz MF, 1,	□ and □ in lipid	□ and □ in NO		MPO (□ or □) depending on		

n et al. (2008)	liver and heart tissues	2, or 3 mT, 4 or 8 h/day for 5 days	peroxidation			exposure condition (duration and intensity) and tissue studied; □ and □ in GSH		
Chen et al. (2014)	Mouse embryonic fibroblast	50-Hz MF, 2 mT, 0.5, 2, 6, 12, 24 h		□ ROS				
Cheun et al. (2007)	Canine kidney MDCK cells	60-Hz MF, 1.4 mT, seconds		MF affected ROS kinetics when H ₂ O ₂ was added to cells.				
Chu et al. (2011)	Mouse cerebellum in vitro	60-Hz MF, 2.3 mT, 3 h	□ lipid peroxidation	□ OH		□ SOD □ GPx		
Chung et al. (2015)	Rat brain in vivo	60-Hz MF, 2.0 mT, 2 or 5 days		□ NO in striatum, thalamus and hippocampus				
Cichon et al. (2017a)	Post-stroke patients	40-Hz, 7 mT for 15 min/day for 4 weeks (5 days a week)				□ SOD and CAT in hemolysates		□ total antioxidant status in plasma; exposed patients showed better improvement in functional and mental status
Cichon et al. (2017b)	Post-stroke patients	40-Hz, 7 mT for 15 min/day for 4 weeks (5 days a week)		□ 3-nitrotyrosine □ nitrate/ nitrite ratio				ELF-EMF promotes recovery of post-stroke patients
Cichon et al. (2018a)	Post-stroke patients, blood mRNA	Rectangular, bipolar waves, 5 mT, 40 Hz, 15 min/ session, 10 sessions in 14 days,						□ CAT, SOD1, SOD2, GPx1, GPx4 mRNA
Cichon et al. (2018b)	Post-stroke patients, blood	Rectangular, bipolar waves, 5 mT, 40 Hz, 15 min/ session, 20 sessions in 20 days,	□ lipid peroxidation □ protein carboxylation					Improved psychophysical abilities of patients

Ciejka et al. (2009)	Sprague-Dawley rats in vivo (plasma)	40-Hz MF, 7 mT, 30 or 60 min per day for 14 days						Repeated 30-min and 60-min exposure increased and decreased plasma antioxidant activity, respectively.
Ciejka et al. (2010)	Sprague-Dawley rats in vivo (muscle)	40-Hz MF, 7 mT, 30 or 60 min per day for 14 days						Both exposures caused an increase in -SH and decrease in proteins in muscle
Ciejka et al. (2011)	Sprague-Dawley rats in vivo (brain)	40-Hz MF, 7 mT, 30 or 60 min per day for 14 days	□ lipid peroxidation in brain of 30-min per day exposed rats					Rats exposed for 60 min per day, 14 days showed increases in -SH and proteins in brain (adaptation).
Ciejka et al. (2014)	Sprague-Dawley rats in vivo (muscle)	40-Hz MF, 7 mT, 30 or 60 min per day for 14 days					GSH	
Coballase-Urrutia et al. (2018)	Restraint Wistar rats	Static MF, 0.8 mT, 30, 60, 240 min/day, 5 days						Attenuated restraint stress-induced increases in NO and MDA and decreases in SOD and GSH
Consales et al. (2018)	SH-SY5Y human neuroblastoma	50-Hz MF, 1 mT, 24, 48 or 72 h	□ O ₂ ^{-•} , H ₂ O ₂					Some ROS produced by mitochondria

	ma cells and mouse primary cortical neurons						a; affected by microRNA (miR-34)
*Consales et al. (2019)	SH-SY5Y human neuroblastoma cells (wild type and two mutants)	50-Hz MF, 1 mT, 24-72 h	∅ O ₂ ⁻ , H ₂ O ₂ ,				∅ iron content and iron-gene expression in a mutant cell-type; ∅ viability and proliferation
Coskun et al. (2009)	Guinea pig in vivo-plasma, brain, and liver	50-Hz MF, 1.5 mT, continuous (C) (4h/day) or intermittent (I) (2 h on/2 h off/2h on) for 4 days	Plasma: I ∅ lipid peroxidation Brain: C, ∅ lipid peroxidation Liver: C, I ∅ lipid peroxidation	Plasma: C, I ∅ NO		MPO: Plasma C ∅ Brain C, I ∅ Liver C, I ∅ GSH: C ∅ I ∅ in brain	
Cui et al. (2012)	C57BL/6 mice in vivo, striatum and hippocampus	50-Hz MF, 1 or 0.1 mT, 4h/day, 12 days	∅ lipid peroxidation in 1 mT group			∅ CAT and ∅ GSH in 1 mT group	∅ Total antioxidant capability in 1 mT group
*de Groot et al. (2014)	Normal and chemically-stressed PC12 cells	50-Hz EMF, 30 min or 48 h, up to 1 mT		No effect on ROS production as measured by H ₂ -DCFDA			
* De Mattei et al. (2003)	Bovine articular cartilage explants	75-Hz EMF 1.3 ms pulses, 2.3 mT peak, 24 h		∅ NO			Pulses enhanced Interleukin-1 β -induced NO production
De Nicola et al. (2006)	U937 cells	Static MF, 6 mT, 2 h; 50-Hz MF, 0.07-0.1 mT, 2 h		∅ ROS		∅ GSH	Decreased apoptosis
Deng B. et al. (2014)	Rat primary cerebral cortical neurons	Electromagnetic pulses (peak 400 KV/m, width 350 ns, 0.5 pps, 1 Hz)	∅ lipid peroxidation			∅ SOD	Decreased cell viability observed, effects antagonized by

								sevoflurane
Deng Y. et al. (2013)	SPF Kunming mouse in vivo, serum and brain	50-Hz MF, 2 mT, 4h/day, 8 weeks	□ lipid peroxidation			□SOD		No interaction with aluminum
Di et al. (2012)	Human preosteoclast FLG29.1 cells	Large gradient high magnetic fields (12 T, -1370 T ² /m; 12 T, 1370 T ² /m), 72 h		□ NO				
*Di Loreto et al. (2009)	Rat cortical neurons	50-Hz MF, 0.1 or 1 mT, 7 days	∅ lipid peroxidation	∅ total ROS		∅ GSH		□ cell viability, □ apoptosis
Dinčić et al. (2018)	Wistar albino rats	Static magnetic field, 1 mT, 50 days	□ lipid peroxidation			□ synaptosomal CAT depending on orientation of static MF		□ ATPase and AchE in synaptosomes
Ding et al. (2004)	Human leukemia HL-60 cells	60-Hz MF, 5 mT, 24 h						Enhanced apoptotic effect of H ₂ O ₂
Djordjevic et al. (2017)	Wistar male rats	50-Hz MF, 10 mT, 7 days, 24 h/day		□ O ₂ ^{-•} and NO; ∅ peroxynitrite (ONOO-) in hypothalamus				
Dornelles et al. (2017)	Human peripheral mononuclear cells with different polymorphism at Val118MnSOD gene	Static magnetic field, 5 mT; 0,1,3,6 h	□ and ∅ lipid peroxidation, □ and □ protein carboxylation	□, □ and ∅ ROS		□, □ and ∅ in SOD1, SOD2, GPX, CAT		Response depended on genetic makeup of the cells
Duan Y. et al. (2013)	ICR mouse, Serum and hippocampus	50-Hz MF, 8 mT, 4 h/day, 28 days	□ lipid peroxidation	□NO	□NOS	□SOD □ CAT □GPx		Effects reversed by lotus seedpot procyanidins
*Duan W. et al.	Mouse spermatocyte-derived	50-Hz EMF, 1, 2, or 3 mT, 5-min on	∅ oxidative DNA base damage					

(2015)	GC-2 cells	10-min off, 24 h						
Duong and Kim (2016)	Human microglial HMO6	50-Hz EMF, 1 mT, 4 h						EMF exposure decreased ROS induced by oxygen-glucose deprivation.
Ehnert et al. (2017)	Human osteoblasts	Pulsed EMF, 16-Hz, 6-282 μ T; 7 min or 7 min/day (>3 days)		Single exposure \square ROS; Repeated exposure \square ROS; mainly $O_2^{\cdot-}$ and H_2O_2		\square GPX3, SOD2, CAT, GSR	Effects of EMF blocked by $O_2^{\cdot-}$ and H_2O_2 scavengers	EMF promotes osteoblast differentiation via free radicals
Emre et al. (2011)	Wistar rat in vivo, liver	Pulsed EMF (0.5 ms rise time, 9.5 ms fall time) EF 0.6 V/m, MF 1.5 mT, each frequency train of 1 Hz, 10 Hz, 20 Hz and 40 Hz was given for 4-min and with 1-min interval between each frequency (together 20 min.); on each day, three exposure cycles performed (1 h), 1 h per day for 30 days	\square lipid peroxidation			\square SOD		No effect on apoptosis, decreased necrosis.
Erdal et al. (2008)	Male and female Wistar rat in vivo, liver	50-Hz MF, 1 mT, 4/day, 45 days	\emptyset lipid peroxidation					Increased 3-nitrotyrosine (oxidative/nitrosative stress) in

								liver of female rats.
Errico Provenzano et al. (2018)	NB4 cells (human acute promyelocytic leukemia)	50 Hz MF, 2 mT, 8, 16, 24 h		□ ROS				
Falone et al. (2007)	Human neuroblastoma cells SH-SY5Y	59-Hz MF, 1 mT, 96-192 h		∅ ROS		∅ SOD, CAT □ GST, GPx 5x □ reduced/ total GSH ratio		Increased cell viability; ∅ cell cycle, apoptosis and DNA damage, but enhanced these effects induced by H ₂ O ₂ .
Falone et al. (2008)	Female Sprague-Dawley rat in vivo, 3- and 19-month old, brain cortex	50-Hz MF, 0.1 mT, 10 days				□ SOD ₂ in young rats; □ catalase and GPx in old rats		□ Glutathione reductase in old and young rats, □ glutathione-S-transferase in old rats: old and young rats responded differently.
Falone et al. (2016)	Human drug-resistant neuroblastoma SK-N-BE(2) cells	72-Hz pulsed EMF, 1.3 ms pulse duration, 2 mT, 15 min, 3 times over 5 days						Pulsed EMF increased MnSOD-based antioxidant protection and reduced ROS production induced by H ₂ O ₂ .
Falone et al. (2017)	SH-SY5Y human neuroblastoma cells	50-Hz MF, 0.1 or 1 mT, 5 and 10 days	□ protein carboxylation and DNA oxidation			□ GPx/ SOD and catalase/ SOD ratios, i.e., increase antioxidating defense; □ GPx activity		Protects cell death by H ₂ O ₂ , □ Nrf2 activity
Feng et al.	Human amniotic	50-Hz MF, 0.4 mT, 5,		□ ROS			MF-induced mitochondri	

(2016a)	epithelial cells	15, 30 or 60 min					al permeability transition blocked by NAC	
Feng et al. (2016b)	Human amniotic epithelial cells	50-Hz MF, 0.1, 0.2, or 0.4 mT, 5, 15, 30, or 60 min		□ ROS				□ total ROS at 0.2 mT and higher, □NADPH oxidase-produced superoxide
Feng et al. (2016c)	Human amniotic epithelial cells	50-Hz MF, 0.2-2 mT, 30, 60, 120 min		□ mitochondrial ROS				□ ROS led to activation of Akt and anti-apoptotic effect
Fernie & Bird (2001)	American kestrel	60-Hz EMF, 30 μT, 10 kV/m, 91 days, 23.5 h/day						Decreased plasma carotenoids
Fiorani et al. (1997)	Rabbit red blood cells	50-Hz MF, 0.2-0.5 mT, 90 min in the presence of an oxygen-generating system (Fe(II)/ascorbate)						Enhanced GSH reduction and hemoglobin oxidation caused by Fe(II)/ascorbate at 0.5 mT
Fitzsimmons et al. (2008)	Human chondrocyte	Pulsed electric field, EF in culture medium 0.2 mV/cm, 30 min		□NO				□cGMP, calcium involved
Frahm et al. (2006)	Mouse bone-marrow derived macrophage	50-Hz MF, 0.05, 0.1, 0.5, 1.0 mT, 45 min		□ROS				
Frahm et al. (2010)	Mouse bone-marrow derived	50-Hz MF, 1.0 mT, 45 min		□ROS				Activated enzymes (NAD(P)H oxidases)

	macrophage							and proteins involved in redox homeostasis
Garip and Akan (2010)	K562 human leukemia cells, normal or treated with H ₂ O ₂	50-Hz EMF 1 mT, 3 h		□ ROS				Decreased and increased apoptosis in untreated and H ₂ O ₂ -treated cells, respectively.
Ghodbane et al. (2011a)	Wistar male rat in vivo, plasma	Static MF, 128 mT, 1 h/day, 5 days	∅ lipid peroxidation			□ GPx		Decreased vitamin A and E levels, effects blocked by selenium
Ghodbane et al. (2011b)	Wistar male rat in vivo, liver, kidney, muscle, brain	Static MF, 128 mT, 1 h/day, 5 days				□ SOD in liver, □ GPx in kidney and muscle, □ GSH in liver		Selenium reversed GPx effect in kidney and muscle
Ghodbane et al. (2014)	Wistar male rat in vivo, plasma	Static MF, 128 mT, 1 h/day, 5 days					Vitamin E blocked static MF effects on blood glucose and liver glycogen	
Ghodbane et al. (2015a)	Wistar male rat in vivo, brain and liver	Static MF, 128 mT, 1 h/day, 5 days	∅ lipid peroxidation in brain and liver			□ CAT in liver	selenium and vitamin E reversed liver catalase effect.	□ apoptosis in liver through a mitochondrial caspase-independent pathway
Ghodbane et al. (2015b)	Wistar male rat in vivo, kidney and	Static MF, 128 mT, 1 h/day, 5	□ lipid peroxidation in kidney			□ CAT in kidney	vitamin E reversed lipid	Selenium reversed □ lipid

	muscle	days					peroxidation effect.	peroxidation and CAT effects in kidney
*Giorgi et al. (2014)	Human neuroblastoma BE(2) cells	Bipolar pulsed square wave MF, 50 Hz, 1 mT, up to 72 h	MF did not affect H ₂ O ₂ -induced DNA double strand break.					
Giorgi et al. (2017)	Human neuroblastoma BE(2) cells	Bipolar pulsed square wave MF, 50 Hz, 1 mT, (average rate of change in MF 3.3 T/s) 24 or 48 h	MF \square DNA methylation with 24 h exposure (not with 48 h)					Oxidative stress (300 μ M H ₂ O ₂) decreased DNA methylation compared to PMF alone
Glinka et al. (2013)	Male Sprague-Dawley rat in vivo, blood serum and liver	40-Hz MF, 10 mT, 30 min / day for 6, 10, or 14 days	\square lipid peroxidation in liver of 6-day exposure			\square SOD-Mn in serum only in 6 day exposure, \square SOD-Mn in liver in 14-day exposure. No effect on SOD-ZnCu \square GPx in serum in 10- and 14-day exposure		\square glutathione s-transferase in liver of 6-day exposure
Glinka et al. (2018)	Mouse fibroblasts	Static magnetic field, 0.1-0.7 T, 72 h	\emptyset lipid peroxidation			\square SOD and GPx		
Gokal et al. (2016)	Wistar rat in vivo, brain and retina	50-Hz EF, 12 kV/m, 1 h/day during prenatal, postnatal, and prenatal + postnatal period	\square lipid peroxidation in brain and retina of exposed animals					Prolonged visual evoked potentials were observed in exposed animals.
Goraca et al. (2010)	Male Wistar rat in vivo, heart and plasma	40-Hz MF, 7 mT, 30 or 60 min/day, 14 days	\square lipid peroxidation in heart in 30 and 60 min/day exposure	\square H ₂ O ₂ in heart in 30 and 60 min/day exposure		\square GSH in heart 60min/day		Total free – SH decreased in heart of 60 min/day, decreased reducing capability in plasma

								of 60 min/day
Güler et al (2008)	Male guinea pig in vivo, liver	50-Hz EF, 12 kV/ m, 8 h/ day, 7 days	□ lipid peroxidation	□ NO		□ SOD □ GPx □ MPO	Blocked by NAC	
*Güler et al (2009a)	Male guinea pig in vivo, plasma	50-Hz EF, 12 kV/ m, 8 h/ day, 7 days	∅ oxidative protein damage					
Güler et al (2009b)	Male guinea pig in vivo, lung	50-Hz EF, 12 kV/ m, 8 h/ day, 7 days	□ protein carboxylation ∅ lipid peroxidation	∅ NO				
Hajipour Verdom et al. (2018)	Human MCF-7 breast cancer cells and HFF normal fibroblasts	Static magnetic field 10 mT, 24 and 48 h		□ ROS		□ GSH in HFF cells		Decreased viability and differentiation in both cell types; Synergistic with doxorubicin
Hajnorouzi et al. (2011)	Maize seedling	Combination of geomagnetic field (47 μT) and perpendicular 10-kHz MF (22 μT), 5 h/ day for 4 days				□ SOD		□ Total antioxidant capacity, faster growth of seedlings, decrease iron increased growth
Hanini et al. (2017)	Mutant Pseudomonas aeruginosa without Mn- and Fe-SOD	Static magnetic field, 200 mT	□ lipid peroxidation			□ SOD, CAT, peroxidases		Wide type bacteria less responsive to the field
*Harakawa et al. (2005)	Sprague-Dawley rat in vivo, plasma	50-Hz EF, 17.5 kV/ m, 15 min/ day, 7 days	∅ lipid peroxidation (□ in oxidatively stressed rats)					No effect on total antioxidant activity
Hashish et al., (2008)	Male Swiss (BALB/ c) mouse in vivo, liver	Static MF (+/- 2.9 μT), or 50-Hz MF 1.4 mT, 30 days	□ lipid peroxidation			□ GSH in ELF-MF exposure only		□ glutathione s-transferase
Henryko	Human	50-Hz MF,	□ lipid peroxidation	□ ROS		□ SOD-1		Effects not

waska et al. (2009)	blood platelet	10 mT, 15 min (sinusoidal, triangular, or rectangular)				□cat alase		wave-shape dependent
*Hong et al. (2012)	Human breast epithelial cells (MCF10A)	60-Hz MF, 1 mT, 4 h		∅ ROS level		∅ SOD ∅ GSH		
Hosseina badi and Khanjani (2019)	Power-plant workers, serum	Chronic ELF-EMF average electric and magnetic field of 4.09 V/m and 16.27 μT						□ SOD, CAT, GPx, and lipid peroxidation on serum correlated with prevalence of musculoskeletal disorders
Höytö et al. (2017)	Human SH-SY5Y neuroblastoma cells	50-Hz MF, 0.1 mT, 24 h		□ cytosolic O ₂ ^{-•} production; □ mitochondrial O ₂ ^{-•} production				
Hu et al., (2016)	3xTG mouse. hippocampus	50-Hz MF, 0.5 mT, 20 h/day for three months		□ ROS				Decreased cognitive deficits, decreased apoptosis; decreased molecules involved in oxidative stress
Jajte et al. (2001)	Rat lymphocyte	50-Hz MF, 7 mT, 3 h	DNA strand breaks				DNA strand breaks induced by MF and FeCl ₂ blocked by melatonin.	
Jajte et al. (2002)	Rat lymphocyte	Static MF, 7 mT, 3 h	□ lipid peroxidation with MF + FeCl ₂					
Jajte et al.	Rat lymphocyte	Static MF, 7 mT, 3 h	□ lipid peroxidation with MF + FeCl ₂				Effect blocked by	

(2003)							melatonin and vitamin E	
Jelenković et al. (2006)	Male Wistar rat in vivo, different brain regions	50-Hz MF, 0.5 mT, 7 days	□ lipid peroxidation in basal forebrain only	□ O ₂ ⁻ □ NO		□ SOD in basal forebrain only		Different brain regions responded differently.
Jeong et al. (2006)	Male ICR mouse in vivo, brain and spinal cord	60-Hz MF, 2 mT, 48 h		□ NO	∅ nNOS, eNOS, iNOS			Hyperalgesia observed, blocked by Ca ²⁺ channel blocker
*Jin et al. (2015)	Human lung epithelial L132 cell	60-Hz MF, 1 or 2 mT, 9 h						MF did not affect H ₂ O ₂ -induced G2/M-arrested or aneuploid cells.
*Jin et al. (2012)	Mouse embryonic fibroblast NIH3T3 and human lung fibroblast WI-38 cells	60-Hz MF, 1 mT, 4 h						MF did not affect H ₂ O ₂ -induced micronucleus formation.
*Jin et al. (2014)	Mouse embryonic fibroblast NIH3T3, human lung fibroblast WI-38, human lung epithelial L132, and human mammary epithelial MCF10A cells	60-Hz MF, 1 mT, 4 or 16 h						MF did not affect H ₂ O ₂ -induced DNA strand breaks.
Jouni et al. (2012)	Broad bean (Vicia faba L.)	Static MF, 15 mT, 8 h/day, 8 days	□ lipid peroxidation			□ SOD □ CAT and peroxidase		
Kamalipoya et al. (2017)	Human cervix cancer	Static MF; 7, 10-16 mT, 24 or 48 h	Synergistic □ lipid peroxidation in cisplatin-treated cancer cells	Synergistic □ ROS in cisplatin-treated cancer cells				Generally, no effect on normal

	(HeLa) Cell, human fibroblasts							fibroblast cells; 10 mT caused highest effects in cancer cells
Kantar Gok et al. (2014)	Male Wistar rat in vivo, brain	50-Hz EF, 12 or 18 kV/m for 2 or 4 weeks, 1 h/day	□ protein carboxylation in 18 kV/m 2 wk and 12 and 18 kV/m 4 wk □ lipid peroxidation in all exposed groups					
Karimi et al. (2019)	Male Wistar rats	50-Hz EMF, 1,100. 500, 2000 μT, 2h/day 60 days	□ lipid peroxidation	□ total thio molecules, □ total oxidant staus		□ total antioxidant activity		Rats showed improved memory retention.
Kavaliers et al. (1998)	Land snail (Cepaea nemoralis) in vivo	60-Hz MF, 0.141 mT, 15 min		□ NO (possible)				MF attenuated opioid-induced analgesia by increasing NO activity
*Kesari et al. (2015)	Human neuroblastoma SH-SY5Y cells	50-Hz MF, 0.1 mT, 24 h	∅ lipid peroxidation	∅ ROS change at 15, 30, and 45 days after exposure				
Kesari et al. (2016)	Human neuroblastoma SH-SY5Y cells and rat C6 glioma cells, cells treated with menadione	50-Hz MF, 10 or 30 μT, 24 h		□ O ₂ ⁻ cytosolic and mitochondrial in C6 cells				Increased micronucleus in SH-SY5Y cells at 30 μT
Khadir et al. (1999)	Human neutrophils simulated by phorbol 12-myristate 13-acetate	60-Hz MF, 22 mT, 10 min		□ O ₂ ⁻				
Kim et al. (2017)	RAW 264.7 macrophage	60-Hz MF, 0.8 mT, up to 20 h		□ NO				Decreased effectiveness of antioxidant

								; increased macrophage activation
*Kimsa-Dudek et al (2018)	Human dermal fibroblast	Static magnetic field, 0.65 T, 24 h		∅ ROS				Changes in antioxidant defense system – related gene expression; Attenuate fluoride-induced changes in antioxidant defense system gene expression
Koh et al. (2008)	Human prostate cancer cells (DU145, PC3, and LNCaP)	60-Hz MF, 1 mT, 6, 24, 48, 72 h		□ H ₂ O ₂			Blocked by NAC	Apoptosis and cell cycle arrest observed.
Koyama et al. (2004)	pTN89 plasmids	60-Hz MF, 5 mT, 4 h						MF potentiated H ₂ O ₂ -induced mutation
Koyama et al. (2008)	Human glioblastoma A172 cell	60-Hz MF, 5 mT, 2, 4, 8, 16, or 24 h						MF potentiated H ₂ O ₂ -induced increase in apurinic/apyrimidinic sites (DNA lesion)
Kthiri et al. (2019)	Saccharomyces cerevisiae (yeast)	Static magnetic field, 250 mT, 6 and 9 h	□ lipid peroxidation and protein carboxylation			□ SOD and CAT □ GP _x after 9 h exposure		Decrease in growth after 6 h and an increase between 6 and 9 h
Kunt et	47 electrical	Mean						□ oxidative

al. (2016)	workers in power transmission facility, serum	working period 15.9 ± 6.72 yrs						stress index (increased total oxidant status, decreased antioxidant status)
Kurzeja et al. (2013)	Mouse fibroblast	Static MF, 0.4, 0.6, and 0.7 T, 4 days						Static MF reduced oxidative stress induced by fluoride ion by normalizing antioxidant enzymes.
Kuzay et al. (2017)	Healthy and diabetic male Wistar rats, testis tissue	50-Hz MF, 8.2 mT, 20 min/day. 5 days/week. 1 month	□ lipid peroxidation	□ NO		□ GSH		
Lai and Singh (1998)	Sprague-Dawley rat in vivo, brain	60-Hz MF, 0.5 mT, 2 h	DNA strand breaks				DNA strand breaks blocked by melatonin and a spin-trap compound.	
Lai and Singh (2004)	Sprague-Dawley rat in vivo, brain	60-Hz MF, 0.01 mT, 24 or 48 h	DNA strand breaks				DNA strand breaks blocked by Trolox and a nitric oxide synthase inhibitor.	Effects blocked by the iron chelator deferiprone
Lai et al. (2016)	Molt-4 human leukemia cells	0.2 Hz pulses, carrier modulated 134 KHz field from radiofrequency ID chip, 1 h					Effect blocked by the spin-trap compound N-tert-butyl-alpha-phenylnitron	Cell death, effect also blocked by the iron-chelator deferoxamine
Lee et al.	Balb/c mouse in	60-Hz MF, 1.2 mT, 3 h	□ lipid peroxidation	∅ O ₂ ⁻		□ SOD		

(2004)	vivo, brain						
*Lee et al. (2012)	Mouse fibroblast NIH3T3	60-Hz MF, 1 mT, 4 h					MF did not affect H ₂ O ₂ -induced cellular transformation
Lee et al. (2010)	Human intervertebral disc cells	60-Hz EMF, 1.8 mT, 72 h					EMF induced DNA synthesis blocked by NMDA, a NO blocker
Lewicka et al. (2015)	Human blood platelet	EMF (1 kHz, 0.5 mT; 50 Hz, 10 mT, 1 kHz, 220 V/m), 30 min	□ lipid peroxidation			□ CAT	
*Li et al. (2015)	Human workers performed inspection near transformers and power lines, plasma	8-h time weighed average magnetic flux intensity 7.3 μT (1.56-26.33 μT), controls 0.07-0.72 μT	∅ lipid peroxidation			∅ SOD ∅ GPx	∅ Total antioxidant capacity, no change in micronucleus frequency
Li et al. (2013)	Male Drosophila melanogaster in vivo	50-Hz EMF, 72 h or long term (312 h), 3 mT					Short term exposure down-regulated CAT gene (endogenous antioxidant enzymes), trend of recovery with long term exposure
Lian et al. (2018)	Yeast (NT64C and SB34)	So Hz-M, 6 mT, 0.5- 24 h		□ ROS at 0.5, 1 ad 2 h		□ SOD at 1 h, □ CAT at 0.5 and 2 h	□ generation and propagation of yeast prions; no ahnge in

								molecular chaperones (several heat-shock proteins)
Liu et al. (2014)	Sprague-Dawley rat cerebellum neurons	50-Hz MF, 1 mT, 1 h						Melatonin (MT) blocked MF-induced Na_v current, MT_2 receptor involved
Liu et al. (2002)	Mouse in vivo, brain and liver	50-Hz EMF, 0.2 or 6 mT, 2 weeks	\square lipid peroxidation, brain and liver			\square GSH in liver		\square decreased total antioxidant capacity in brain and liver, decreased cell membrane fluidity, synergism with lead
Luo et al. (2016)	ICR mouse Blood and cerebral cortex	50-Hz, 2-10 mT, 4 h/days. 28 days	\square lipid peroxidation in serum and cerebral cortex			\square SOD, \square CAT, \square glutathione reductase, \square GSH-Px, and glutathione-s-transferase in serum and cerebral cortex		
Luo et al. (2019)	Sitobion avenae Fabricius (a herbivorous insect)	High-voltage electric field (HVEF); 2, 4, or 6 kV/cm; 20, 40, or 60 min; assayed up to 21 generations				\square SOD, \square CAT, \square peroxidase over multiple generations		Exposed insects have higher Co2 production rate
Lupke et al. (2004)	Human umbilical cord blood derived monocyte and human mono Mac 6 cells	50-Hz MF, 1 mT, 45 min			\square total ROS, $\square \text{O}_2^{\cdot-}$			Mono Mac 6 cells more sensitive, activation of NADPH oxidase not NADH oxidase.
Luukkonen et al.	Human SH-SY5Y	50-Hz MF, 0.1 mT, 24 h			\square ROS, $\square \text{H}_2\text{O}_2$ in mitochondria			interacts with

(2014)	neuroblastoma cell							menadione; effects observed days after exposure
Malhמודi nasab et al. (2016)	Human MCF-7 cells	50-Hz EMF, 0.25 and 0.5 mT; 5-min on/ 5-min off; 15-min on/ 15-min-off, or 30 min continuously; total exposure time 30 min						Changes in mRNA levels of 7 antioxidant genes
Malhמודi nasab and Saadat (2018a)	Human SH-SY5Y and MCF-7 cells	50-Hz EMF, 0.5 mT, 15 min on/ 15 min off						Up-regulation of antioxidant genes and protection of Cisplatin cytotoxicity in SH-SY5Y cells, but not MCF-7 cells
Malhמודi nasab and Saadat (2018b)	Human SHSY5Y cells	50-Hz EMFm 0.5 mT, 15 min on/ 15 min off or 30 min continuously						Changes in antioxidant gene NQO1 and NQO2
Maliszewska et al. (2018)	American cockroach (Periplaneta Americana L)	50-Hz EMF, 7 mT, 24, or 72 h or 7 days	□ lipid peroxidation			□ GSH		
Manikonda et al. (2014)	Male Wistar rat in vivo, brain (hippocampus, cerebellum and cortex)	50-Hz MF, 0.05 and 0.1 mT, 90 days	□ lipid peroxidation	□ ROS		□ SOD □ GSH/ GSSG ratio		Larger response at 0.1 mT
Mannerling et al. (2010)	Human leukemia cell K562	50-Hz MF, 0.025-0.1 mT, 1 h		□ O ₂ ⁻			Melatonin blocked MF-induced	

							HSP70	
*Markkane n et al. (2010)	Murine L929 fibroblast	50-Hz MF, 0.1-0.3 mT, 1 h						Did not affect ROS production induced by UV.
Martinez et al. (2016)	Human neuroblastom a NB69 cells	50-Hz MF, 0.1 mT, 3-h on/3-h off for 24, 42, or 63 h, or continuously for 15-120 min					MF-induced MAPK-p38 and ERK1/2 activation blocked by NAC	
Martine z- Samano et al. (2010)	Male Wistar rat in vivo, plasma, liver, kidney and heart	60-Hz EMF, 2.4 mT, 2 h	∅ lipid peoxidation				∅ SOD in plasma of MF and restrained rats ∅ CAT ∅ GSH in heart	Interacts with restraint stress
Martine z- Samano et al. (2012)	Male Wistar rat in vivo, brain	60-Hz EMF, 2.4 mT, 2 h					∅ SOD ∅ CAT ∅ GSH	Interacts with restraint stress
Martine z- Samano et al. (2018)	Male Wistar rat in vivo, brain	60-Hz EMF, 2.4 mT, 2 h/day, 21 days	∅ lipid peroxidation in cortex and cerebellum					∅ plasma corticoster one
Martino (2011)	Human umbilical vein endothelial cell	Static MF, 0.12 and 0.03 mT (compared to 0.2-1 μ T), 2 days						Increased cell proliferation attenuated by SOD
Martino and Castello (2011)	Human fibrosarcom a HT1080, pancreatic AsPC-1 cancer cells, and bovine pulmonary artery endothelial cells	Static MF, geomagnetic field (45-60 μ T) or shielded field (0.2-2 μ T), 24 h		∅ H ₂ O ₂ in shielded samples compared to geomagnetic field				MnTBAP (a ROS scavenger) inhibited MF effect.

Medina-Fernandez et al. (2017)	Ale Dark Agouti rat; experimental model of multiple sclerosis induced by myelin oligodendrocyte glycoprotein (MOG), brain and spinal cord	Transcranial magnetic stimulation (TMS), 60-Hz, 0.7 mT, 2 h/day, 5 days a week, 3 weeks	TMS reversed protein carboxylation and lipid peroxidation induced by MOG			□ GSH		Decreased cell death and motor deficit induced by MOG
Medina-Fernandez et al. (2018)	Ale Dark Agouti rat; experimental autoimmune encephalomyelitis induced by myelin oligodendrocyte glycoprotein (MOG), brain, spinal cord and blood	Transcranial magnetic stimulation (TMS), 60-Hz, 0.7 mT, 2 h/day, 5 days a week, 3 weeks	TMS reversed protein carboxylation and lipid peroxidation induced by MOG			□ GSH		TMS reversed tail and limb paralysis induced by MOG
Merle et al. (2019)	Human SH-SY5Y neuroblastoma cells	50-Hz MF, 1 mT, 24 h		□ O ₂ ^{-•} and H ₂ O ₂				Effects involved NADPH-oxidase on plasma membrane

Miao et al. (2017)	Male BALB/c mice, in vivo, testicle	Electromagnetic pulse, 200 kV/m, pulse edge 25 nms pulse width 15 ns, 0.1 Hz, 40 pulses/day, 5 days/week, 4 weeks				□ Testicular antioxidant capacity at 28 and 60 days after exposure		□ spermatozoa formation
Miliša et al. (2017)	Euglena viridis and Paramecium caudatum	50-Hz EF, 2.5, 5.0, 9.3 and 13.6 kV/m, 24 h		□ O ₂ ^{•-} and H ₂ O ₂		□ SOD		
*Missiha et al. (2015)	Flavin-dependent redox enzymes	Static MF, 10-160 mT, seconds						MF did not change enzyme kinetics. Radical pair not a mechanism of redox reaction with static MF.
Mohammadi et al. (2018)	Tobacco cells	Static Magnetic field, 0.2 mT, 24 h		□ H ₂ O ₂ , □ NO				Delayed G1-S transition, increased cyclic nucleotides
Morabito et al. (2010a)	Rat pheochromocytoma PC-12 cell	50-Hz MF. 0.1 or 1 mT, 30 min or 7 days		□ ROS in 30 min exposure at 1 mT.		□ CAT in 0.1 and 1 mT 30-min exposure, □ catalase in 1 mT 7-day exposure		All effects were observed in undifferentiated and not in differentiated cells. Calcium probably involved.
Morabito et al. (2010b)	Undifferentiated C2C12 myoblast	50-Hz MF. 0.1 or 1 mT, 30 min		∅ O ₂ ^{•-} □ H ₂ O ₂ in 1 mT exposure		□ CAT and GPx	NAC attenuated free radical increase by MF	Calcium probably involved.
Naarala	Rat glioma	Nearly		□ cytosolic O ₂ ^{•-} in vertical				Cell

et al. (2017)	C6 cells	vertical 33 μ T static MF plus a horizontal or a vertical 50-Hz 30 μ T MF, 2 h		static field plus horizontal 50-Hz MF (but not vertical 50-Hz MF); Mitochondrial $O_2^{\cdot-}$ not affected				viability not affected.
*Nakayama et al. (2016)	Mouse macrophage (RAW 264) with or without LPS stimulation	50-Hz MF, 0.5 mT, 24 h		\emptyset NO				
Noda et al. (2000)	Rat brain cerebellum tissues	Pulsed DC MF, 0.1 mT, 1 h			\square NOS			No effect from pulsed DC at 0.3 and 0.6 mT, 60 Hz (0.1 mT), and DC (3 or 20 mT) MF, no effect in hippocampus, cortex, medulla oblongata, hypothalamus, striatum, and midbrain.
Osera et al. (2011)	Human neuroblastoma SH-SY5Y cells	72-Hz pulsed EMF, 1.3 ms pulse duration, 2 mT, 72 h				\square SOD-1		Increased quiescent cells
Osera et al. (2015)	Human neuroblastoma SH-SY5Y cells	72-Hz pulsed EMF, 1.3 ms pulse duration, 2 mT, 10, 15, or 30 min for 4 times over 7 days, or 72 h				\square Mn-SOD		Interacts with H_2O_2 . Pulsed EMF prevented H_2O_2 - induced decrease in cell number and protein expression

								(HSP70). No effect on U937 cells
Pakhomova et al. (2012)	Jurket cells	Nanosecond pulsed electric field (300 ns, 1-12 kV/cm)		□ ROS proportional to pulse number				
Pandir and Sahingoz (2014)	Moth Ephestia kuehniella larvae	Static MF, 1.4 T; 3, 6, 12, 24, 48, or 72 h	□ lipid peroxidation			Exposure-time dependent □ SOD, CAT, GPx and GST		
Park et al. (2013)	Human bone marrow mesenchymal stem cells	50-Hz EMF, 1 mT, 90 min		□ ROS			Blocked by NAC	
Patruno et al. (2010)	Human epidermal keratinocyte cell HaCaT	50-Hz MF, 1 mT, 3, 18, 48 h		□ O ₂ ⁻ □ NO	□ iNOS and eNOS	□ CAT		Increased cell proliferation.
Patruno et al. (2011)	Human epidermal keratinocyte cell HaCaT and acute myeloid leukemia THP-1 cell	50-Hz MF, 1 mT, 24 h			□ iNOS activity	□ CAT activity		
Patruno et al. (2012)	Human acute myeloid leukemia THP-1 cell	50-Hz MF, 1 mT, 24 h		□ O ₂ ⁻	□ iNOS	□ SOD □ CAT		
Patruno et al. (2015)	Human erythro-leukemic K562 cell	50-Hz MF, 1 mT, 24 h			□ iNOS reaction velocity	□ CAT activity		
Pilla (2012)	Human dopaminergic MN9D cells and fibroblasts	Pulsed radiofrequency signal, 2 Hz, with 127.2 MHz carrier; 2.5 μT, 15 min		□ NO				May involve activation of calcium/calmodulin nitric oxide synthase (cNOS)
Politanski et al.	C57BL/g mouse in	Static MF, 5 mT, 2 h,	□ lipid peroxidation in MF + noise'			□ SOD in MF, noise, and 'MF + noise',		MF interacted

(2010)	vivo, cochlear	repeated over 14 days (also exposed to noise once)				□ CAT activity in MF, noise, and 'MF + noise'		with noise
Poniedzialek et al. (2013a)	Human neutrophil	EMF tuned to calcium ion cyclotron resonance frequency (up to 60 μ T)		□ ROS in unstimulated cells, □ in phorbol 12-myristate 13-acetate stimulated cells				
Poniedzialek et al. (2013b)	Human neutrophil	Gradient static MF, maximum value 60 mT, 15, 30 or 45 min		□ ROS in 15-min exposure, □ in 45-min exposure in both unstimulated and phorbol 12-myristate 13-acetate stimulated cells, effect depended on whether samples were placed close to south or north pole of magnet.				
Pooam et al. (2017)	Human macrophage RAW264	50 Hz MF, 0.1 or 0.5 mT, 1, 17 or 24 h		□ O ₂ ⁻				
Potenza et al. (2010)	Human umbilical vein endothelial cells	Static MF, 300 mT, 4, 24, 48, and 72 h		□ ROS only at 4-h exposure coincided with DNA damage				
Rageh et al. (2012)	10 -day old rat in vivo, brain	50-Hz MF, 0.5 mT, 30 days (24 h/day)	□ lipid peroxidation			□ SOD □ GSH		
Raggi et al. (2008)	Human blood sample	Magnetic therapy device based on ion cyclotron resonance	□ lipid peroxidation immediately and one month after exposure					
Rajabbeigi et al. (2013)	Parsley cell	Static MF, 30 mT, 6 or 12 h				□ CAT with MF □ CAT with 'MF + iron'		□ ascorbate peroxidase
Rauš Balind et al. (2014)	Gerbil subjected to 10-min global cerebral ischemia in	50-Hz MF, 0.5 mT, 7 days						MF decreased oxidative stress induced by ischemia

	vivo, brain (forebrain, striatum and hippocampus)							(NO, SOD, MDA, O ₂ ⁻)
Reale et al. (2006)	Human blood monocytes	50-Hz EMF, 1 mT, overnight			□ iNOS			
Reale et al. (2014)	Human neuroblastoma cell SH-SY5Y	50-Hz MF, 1 mT, 1, 3, 6 or 24 h		□ O ₂ ⁻	□ NOS, peaked at 1 h	□ CAT		MF enhanced oxidative effects of H ₂ O ₂ (□cat alase, □ O ₂ ⁻).
Regoli et al. (2005)	Snail Helix aspersa in vivo, digestive gland	50-Hz MF, 0.5, 2.5, 10 and 50 μT, 10 days in lab; 2.88 and 0.75 μT for 10, 20, 40, 60 days in field	Lab: ∅ lipid peroxidation Field: □ in 2.88 μT more than 10 days and 0.75 μT more than 20 days			Lab: □ CAT in 50 μT 10 days Field: □ CAT in 2.88 μT more than 10 days and 0.75 μT more than 40 days Lab: ∅ GSH, □ Glutathione reductase Field: □ glutathione reductase		Total oxyl-radical scavenger capacity: Lab: □OH and □OO; Field: □OH and □OO
Rollwitz et al. (2004)	Mouse bone marrow-derived promonocytes and macrophage	50-Hz MF, 1 mT, 45 min-24 h		□ ROS, □ O ₂ ⁻				NADH-oxidase (not NADPH pathway) involved.
*Romeo et al. (2016)	Human fetal lung fibroblasts (MRC-5)	Static MFm 370 mT, 1 h/day for 4 days		∅ ROS				∅ viability, DNA strand breaks, and apoptosis
Roy et al. (1995)	Phorbol 12-myristate 13-acetate-stimulated rat neutrophil	60-Hz MF, 0.1 mT		□ ROS				
Sadeghipour et al. (2012)	Human breast carcinoma cell (T47D)	100 and 217 Hz pulsed EMF, 0.1 mT, 24-72 h		□ ROS in 217 Hz 72-h, not in 100 Hz exposure				
Sahebjam et al.	Cultured tobacco	Static MF, 10 and 30 mT,	□ lipid peroxidation			□ SOD □ CAT and ascorbate		

(2007)	cell	5 h/day, 5 days				peroxidase		
Salunke et al. (2014)	Swiss albino mouse in vivo, brain	50-Hz MF, 1 mT, 8 h/day for 7, 30, 60, 90 and 120 days		□ NO in cortex, hippocampus, and hypothalamus				
Seif et al. (2018)	Male Wistar rats, in vivo blood	50-Hz EMF, 0.7 mT, 2 h/day. 1 month	□ plasma protein carboxylation, methemoglobin and hemichrome			□ plasma anti-oxidation capacity		
Seifirad et al. (2014)	Male Wistar rat in vivo, serum	60-Hz MF, 0.5 mT, 4 h or 4 h/day 14 days	□ lipid peroxidation immediately after and at 72 h after chronic exposure, ∅ acute exposure					Total antioxidant activity: □ immediately after acute exposure (not at 3 days post-exposure), □ immediately and 3 days after chronic exposure.
Selaković et al. (2013)	Male gerbils 3- and 10-month old in vivo, Forebrain cortex, striatum hippocampus, and cerebellum	50-Hz MF, 0.5, 0.25 and 0.1 mT, 7 days	□ lipid peroxidation	□ O ₂ ⁻ □ NO		□ SOD		Dose-response observed, effects smaller and recovered faster in 3-month than in 10-month old animals.
Sharifian et al. (2009)	Human welders occupational exposure, serum and red blood cells	50-Hz EMF, 8.8-84 μT, 20-133 V/m, 40 h/week (6 days/week)				□ SOD □ GPX		∅ Total serum antioxidant status, a significant negative correlation between SOD/ GPX and MF intensity was observed.

Sherrard et al. (2018)	Insect sf21 cells, human embryonic kidney cells, mouse embryonic fibroblasts	Pulsed EMF. 10 Hz, peak intensity 2 mT, 15 min; with blue light		□ ROS				Effects involved cryptochromes
Shine et al (2012)	Soybean seeds	Static MF 150 and 200 mT, 1 h		□ O ₂ ⁻ , OH, H ₂ O ₂ ,		□ SOD & ascorbate peroxidase		
Shokrolahi et al. (2018)	Soybean plants	Static MF, 20 and 30 mT, 5 days, 5 h/day		At 20 mT, □ H ₂ O ₂ ;; at 30 mT, □ H ₂ O ₂ ,		At 20 mT, □ CAT ₁ ;; at 30 mT, □ CAT ₁ ,		At 20 mT, □ gene expression of Fe transporter, ferrous content,, and gene expression and content of ferritin;; 30 mT produced the opposite effects of these parameters
Simko et al. (2001)	Mouse bone marrow-derived macrophage	50-Hz MF, 0.5-1.5 mT, 45 min		□ O ₂ ⁻				Increased phagocytic activity.
Sirmatel et al. (2007a)	Male human blood	1,5 T static MF from a MRI machine, 30 min						□ total antioxidant capacity; □ total oxidant status and oxidative stress index
Sirmatel et al. (2007b)	Male human blood	1,5 T static MF from a MRI machine,		□ NO (based on nitrite and nitrate levels)				

		30 min						
Solek et al. (2017)	Mouse spermatogenic cell lines	2, 50, 120 Hz pulsed (1 sec on/ 1 sec off) and continuous-wave EMF, 2.5-8 mT, 2 h		□ O ₂ ^{•-} □ NO				Cell cycle arrest and apoptosis observed
Song et al. (2018)	Human cervical cancer cells (HeLa) and lung fibroblasts (IMR-90)	60-Hz EMF 1-10 mT, up to 72 h		□ ROS				Increased cell proliferation
Sullivan et al. (2011)	Various human cell lines	Static MF, 35-120 mT		Static MF □ ROS at 18 h (not at 5 days) of exposure in fetal lung (WI38) cells.				Effects observed in some cell types and not in others.
Sun et al. (2015)	Preosteoclast cell line RAW264.7	Large gradient high magnetic fields (12 T, -1370 T ² /m; 12 T, 1370 T ² /m), 48 h		□ NO				
Sun L. et al. (2018)	Human amnion epithelial cells	50-Hz MF, 0.4 mT, 15 min		□ ROS			Inhibited by N-acetyl-l-cysteine and pyrrolidine dithiocarbamate	Increase in free radicals correlated to clustering of cell surface epidermal growth factor receptor
Sun Y. et al. (2018)	Caenorhabditis elegans	50-Hz EMF, 3 mT. exposed from egg to fourth larva stage		□ ROS				
Tang et al. (2016)	Human Jurket cell and	7.5 Hz MF, 0.4 T, 2 h		□ ROS		□ Total anti-oxidant activity, Ø SOD, Ø CAT		Disruption of tricarboxyly

	stimulated mouse primary T cell							c acid cycle enzymes PGE2 and formation
Tasset et al. (2012)	Male Wistar rat in vivo, brain	60-Hz MF, 0.7 mT, 2 h in the morning and 2 h in the afternoon for 21 days (applied to the head)	Ø DNA oxidative damage Ø lipid peroxidation			Ø GSH □ GSSG		MF reversed 3-nitropropionic acid induced oxidative stress.
Tayefi et al. (2010)	Wistar rat pup in vivo, myocardium	50-Hz MF, 3 mT, 4/h per day during gestation and to 20 day postnatal	□ lipid peroxidation			□ SOD		
Todorovic et al. (2012)	Eggs of Baculum extrudentatum (insert also known as Vietnamese walking stick)	Static MF, 50 mT; 50-Hz MF, 6 mT; exposed until completion of embryonic development				□ SOD and CAT Ø GSH		
Todorovic et al. (2019)	One-month old Blaptica dubia (cockroach) nymphs; gut assayed	110 mT Static MF or 10 mT 50-Hz MF, for 5 months				□ SOD and CAT; Ø GSH; □ GST and glutathione reductase		MF exposure decreased gut mass of developing cockroach
Túnez et al. (2006)	Male Wistar rat in vivo, striatum	60-Hz MF, 0.7 mT, 2 h in the morning and 2 h in the afternoon for 4 days (applied to the head)						MF itself had no effect on different oxidative parameters, but reduced 3-nitropropionic acid induced oxidative and

								nitrosative stress.
*Türközer et al. (2008)	Guinea pig in vivo, brain	50-Hz EF, 2, 2.5, 3, 3.5, 4, 4.5, 5 V/m, 8 h/day, 3 days	∅ lipid peroxidation			∅ SOD ∅ CAT and GPx		
Van Huizen et al. (2019)	Schmidtea mediterranea (planarian), regeneration after amputation	Static magnetic field; 100-400 μ T and 500 μ T; 12, 24 or 48 h		□ ROS after 100-400 μ T and □ ROS after 500 μ T exposure				Reduced blastema (regrowth) size at 100-400 μ T, increased at 500 μ T; ROS altered stem cell proliferation and differentiation depending on field intensity, inhibiting SOD pharmacologically reversed decreased regeneration effect of 200 μ T
*Vannoni et al. (2012)	Human osteoarthritic chondrocyte	100-Hz EMF and a field containing various frequencies		∅ ROS		∅ GSH		
Vignola et al. (2012)	Female Wistar rat with drug-induced myopathy, in vivo, muscle	Pulsed EMF, 50-Hz carrier frequency, 20 mT, 30 min/day, 8 days, assayed 8 days after exposure		□ NO		□ SOD		Pulsed EMF caused muscle recovery.

*Villarini et al. (2017)	SH-SY5Y5 and SK-N-BE-2 human neuroblastoma cells	50-Hz MF; 0.01, 0.1, or 1 mT; 1 h continuously or 5 h intermittently	∅ DNA damage			∅ GSH/ GSSG ratio		
Wang et al. (2019)	11 cancer and normal cell lines	Static magnetic field and 50- and 120-Hz MF, 6 mT, 2, 4, or 6 h		ROS measured in 4 cell lines after 2 h exposure. □ and □ ROS observed depending on cell line and field				No change in ATP levels, □ and □ in mitochondrial membrane potential depending on cell type
Wartenberg et al. (2008)	Oral mucosa cancer cell (UM-SCC-14-C)	DC EF, 4 V/m, 24 h				□ Cu/ Zn SOD ∅ CAT □ GSH	Effects blocked by NAC.	Increased apoptosis and decreased cell proliferation.
Wolf et al. (2005)	HL-60 leukemia cells, Rat-1 fibroblast, WI-38 diploid fibroblast	50-Hz EMF, 0.5-1 mT, 24-72 h	□ DNA oxidative damage	□ ROS in Rat-1 fibroblast			Effect blocked by alpha-tocopherol.	Dose-dependent increase in cell proliferation observed.
Wu et al. (2016)	Male mice, liver	Static E-field, 9.2-21.85 kV/m, 2.3-15.4 kV/m, and 0 kV/m, 35 days	∅ lipid peroxidation			□ SOD		No effect on glutathione transferase and glutathione peroxidase
Yang and Ye (2015)	Human osteosarcoma MG-63 cells	50-Hz EMF; 1 mT; 1,2 or 3 h		□ ROS			Blocked by N-acetylcysteine	□ viability and cell growth; □ apoptosis
Yang et al. (2016)	Sprague-Dawley rats and isolated	EMP, 200 kV/m, 200 pulses; assayed at 1,		□ NO in cerebral cortex of rats and microglial cells, effect returned to normal at 24 h				

	microglial cells	6, 12 and 24 h after exposure						
Yin et al. (2016)	Primary cultured rat hippocampal neurons	50-Hz MF, 8 mT, 90 min	□ lipid peroxidation	□ ROS		□ SOD		
Yokus et al. (2005)	Female Wistar rat in vivo, leukocytes and plasma	50-Hz MF, 0.97 mT, 3 h/day for 50 or 100 days	□ DNA oxidative damage □ lipid peroxidation					Larger effects with longer exposure.
Yokus et al. (2008)	Male Sprague-Dawley rat in vivo, leukocytes	50-Hz MF, 0.1 and 0.5 mT, 2 h/day for 10 months	□ different forms of oxidative DNA damage in 0.1 mT group					
*Yoon et al. (2014)	Human lung fibroblast W138 and human lung epithelial L132 cells	60-Hz MF, 1 or 2 mT, 6 h						MF did not enhance H ₂ O ₂ -induced double strand DNA breaks. (MF potentiated infra-red induced breaks).
*Yoshikawa et al. (2000)	Male BALB/C mouse injected with lipopolysaccharide (LPS) in vivo, liver	60-Hz MF, 0.1 mT, 5.5 h						MF did not induce NO generation, but enhanced LPS-induced NO generation.
Zeng et al. (2011)	Male Sprague-Dawley rats	EMP, 100 kV/m, 50 pps 2.5-2.8 ns width, total pulses 4x 10 ⁵	□ lipid peroxidation in testes			□ total SOD and manganese-SOD in testes		Changes in ultrastructures of testes
Zeng et al. (2017)	Hippocampal neurons from embryonic Sprague-Dawley rats	50-Hz MF, 2 mT; acute; 30 min, 8 h or 24 h on DIV (days in vitro) 7 or 14; repeated: 30 min or 8 h		□ ROS after repeated exposure				□ cell viability; □ expression of NADPH oxidase (subunit Nox2),

		DIV1-7 or DIV 7-14						responsible for ROS production
Zhang et al. (2016)	Canton Special and W1118 flies	50-Hz MFn 30 mT, 12 h						Acted synergistically with heat on oxidative stress and induction of heat shock proteins, effects depends on species and sex
Zhang et al. (2017)	Workers occupationally exposed to EMF in a power plant	>20 yrs	<input type="checkbox"/> DNA oxidative damage (8-OHdG measured in plasma)				Effects reversed by resveratrol (500 mg twice daily, 12 months)	Exposed subjects showed reduced inflammatory biomarkers
Zhang et al. (2018)	RAW 264.7 bone monocytes	Static MF, 500 nT, 0.2 T, 16 T; 12 h to 4 days		<input type="checkbox"/> NO (16 T) <input type="checkbox"/> NO (500 nT and 0.2 T)	<input type="checkbox"/> NOS (16 T) <input type="checkbox"/> NOS (500 nT and 0.2 T)			NO mediates SMF effects on osteoclast formation; effect depends on intensity of MF
Zhao et al. (2011)	Human-hamster hybrid(A9L)), mitochondria-deficient (p(O)A(L)) cells, double-strand break repair-deficient (XRS-5) cells	Static MF, 8.5 T, 3 h		<input type="checkbox"/> ROS				

Zwirska-Korczala et al. (2004)	Murine squamous carcinoma AT478 cell	Mixture of frequencies up to 400 Hz, MF, 0.11 mT, 16 min, assayed 24 and 72 h after exposure	□lipid peroxidation			□ MnSOD and Cu/ ZnSOD ∅ GPx	Effects attenuated by melatonin	
Zmylony et al. (2004a)	Rat lymphocytes stimulated by FeCl ₂	50-HZ MF, 20, 40, or 200 μT, 5 or 60 min		□ ROS in Fe and 40 μT MF exposed cells (AC MF has to be directed along the earth's static MF).				
Zmylony et al. (2004b)	Rat lymphocytes	50-HZ MF, 40 μT, 5 or 60 min	MF enhanced DNA damage caused by ultraviolet radiation (UVA). (UVA damages DNA via free radicals.)					

Literature list (E= 213 (89%); NE= 27 (11%)) (E= paper reported effect; NE= paper reported no significant effect).

(VT = in vitro; VO= in vivo; HU= human study; CE = long-term/repeated exposure; AE= acute exposure; LI = low intensity; IFR= increase free radical; DFR= decrease free radical; IOD = increase oxidative damages; DOD = decrease oxidative damages; IAO =increase antioxidant activity; DAO= decrease antioxidant activity; AO= effect of antioxidant/free radical scavenger; IX= interaction with other factor; MC= mechanism)

(E) (VT, AE, IFR, IAO) Akan Z, Aksu B, Tulunay A, Bilsel S, Inhan-Garip A. Extremely low-frequency electromagnetic fields affect the immune response of monocyte-derived macrophages to pathogens. *Bioelectromagnetics*. 31(8):603-612, 2010.

(E) (VO, CE, DFR) Akdag MZ, Bilgin MH, Dasdag S, Tumer C. Alteration of nitric oxide production in rats exposed to a prolonged, extremely low-frequency magnetic field. *Electromagn Biol Med*. 26(2):99-106, 2007.

(E) (VO, CE, DAO) Akdag MZ, Dasdag S, Ulukaya E, Uzunlar AK, Kurt MA, Taşkın A. Effects of extremely low-frequency magnetic field on caspase activities and oxidative stress values in rat brain. *Biol Trace Elem Res.* 138(1):238-249, 2010.

(E) (VO, CE, IOD) Akdag MZ, Dasdag S, Cakir DU, Yokus B, Kizil G, Kizil M. Do 100- and 500- μ T ELF magnetic fields alter beta-amyloid protein, protein carbonyl and malondialdehyde in rat brains? *Electromagn Biol Med.* 32(3):363-372, 2013a.

(NE) (VO, CE) Akdag MZ, Dasdag S, Uzunlar AK, Ulukaya E, Oral AY, Celik N, Akşen F. Can safe and long-term exposure to extremely low frequency (50 Hz) magnetic fields affect apoptosis, reproduction, and oxidative stress? *Int J Radiat Biol.* 89(12):1053-1060, 2013b.

(E) (VO, CE, IOD) Akpınar D, Ozturk N, Ozen S, Agar A, Yargicoglu P. The effect of different strengths of extremely low-frequency electric fields on antioxidant status, lipid peroxidation, and visual evoked potentials. *Electromagn Biol Med.* 31(4):436-448, 2012.

(E) (VO, CE, DOD, IOD) Akpınar D, Gok DK, Hidisoglu E, Aslan M, Ozen S, Agar A, Yargicoglu P. Effects of pre- and postnatal exposure to extremely low-frequency electric fields on mismatch negativity component of the auditory event-related potentials: Relation to oxidative stress. *Electromagn Biol Med.* 35(3):245-259, 2016.

(E)(VO, CE, IOD) Aksen F, Akdag MZ, Ketani A, Yokus B, Kaya A, Dasdag S. Effect of 50-Hz 1-mT magnetic field on the uterus and ovaries of rats (electron microscopy evaluation). *Med Sci Monit.* 12(6):BR215-220, 2006.

(NE) (VO, CE, AO) Alcaraz M, Olmos E, Alcaraz-Saura M, Achel DG, Castillo J. Effect of long-term 50 Hz magnetic field exposure on the micronucleated polychromatic erythrocytes of mice. *Electromagn Biol Med.* 33(1):51-57, 2014.

(E) (VO, CE, DAO, IOD) Amara S, Douki T, Garel C, Favier A, Sakly M, Rhouma KB, Abdelmelek H. Effects of static magnetic field exposure on antioxidative enzymes activity and DNA in rat brain. *Gen Physiol Biophys.* 28(3):260-265, 2009.

(E) (VO, CE, DAO, IOD, IX) Amara S, Douki T, Garrel C, Favier A, Ben Rhouma K, Sakly M, Abdelmelek H. Effects of static magnetic field and cadmium on oxidative stress and DNA damage in rat cortex brain and hippocampus. *Toxicol Ind Health*. 27(2):99-106, 2011.

(E) (VO, AE, CE, IAO) Ansari AM, Farzampour S, Sadr A, Shekarchi B, Majidzadeh-A K. Effects of short term and long term extremely low frequency magnetic field on depressive disorder in mice: Involvement of nitric oxide pathway. *Life Sci*. 146:52-57, 2016.

(E) (VO, AE, IAO) Asghar T, Jamil Y, Iqbal M, Zia-Ul-Haq, Abbas M. Laser light and magnetic field stimulation effect on biochemical, enzymes activities and chlorophyll contents in soybean seeds and seedlings during early growth stages. *J Photochem Photobiol B*. 165:283-290, 2016.

(E) (VT, CE, IFR) Ayşe IG, Zafer A, Sule O, Işıl IT, Kalkan T. Differentiation of K562 cells under ELF-EMF applied at different time courses. *Electromagn Biol Med*. 29(3):122-130, 2010.

Barnes F, Greenebaum B. Role of radical pairs and feedback in weak radio frequency field effects on biological systems. *Environ Res*. 163:165-170, 2018. (Review)

(E) (VT, AC, IX) Bawin SM, Satmary WM, Jones RA, Adey WR, Zimmerman G. Extremely-low-frequency magnetic fields disrupt rhythmic slow activity in rat hippocampal slices. *Bioelectromagnetics*. 17(5):388-395, 1996.

(E) (VO, CE, IOD) Bediz CS, Baltaci AK, Mogulkoc R, Oztekin E. Zinc supplementation ameliorates electromagnetic field-induced lipid peroxidation in the rat brain. *Tohoku J Exp Med*. 208(2):133-140, 2006.

(E) (VT, AE, IFR, DFR, MC) Belova NA, Potselueva MM, Skrebnitskaia LK, Znobishcheva AV, Lednev VV. The influence of weak magnetic fields on the production of the reactive oxygen species in peritoneal neutrophils in mice. *Biophysics (Biofizika)*. 55(4):586-591, 2010.

(E) (VT, AE, IOD, IX) Benassi B, Filomeni G, Montagna C, Merla C, Lopresto V, Pinto R, Marino C, Consales C. Extremely low frequency magnetic field (ELF-MF) exposure sensitizes SH-SY5Y cells to the pro-Parkinson's Disease toxin MPP. *Mol Neurobiol*. 53(6):4247-4260, 2016.

(E) (VO, AE, DAO) Berteza CM, Narayana R, Agliassa C, Rodgers CT, Maffei ME. Geomagnetic Field (Gmf) and Plant Evolution: Investigating the Effects of Gmf Reversal on Arabidopsis thaliana Development and Gene Expression. *J Vis Exp.* (105), 2015.

Brocklehurst B, McLauchlan KA. Free radical mechanism for the effects of environmental electromagnetic fields on biological systems. *Int J Radiat Biol.* 69(1):3-24, 1996. (Review)

(E) (VT, AE, IOD) Buczyński A, Pacholski K, Dziedziczak-Buczyńska M, Henrykowska G, Jerominko A. The assessment of oxygen metabolism selected parameters of blood platelets exposed to low frequency magnetic radiation in cars--in vitro studies. *Rocz Akad Med Białymst.* 50 Suppl 1:23-25, 2005.

(E) (VO, CE, LI, DAO, IAO) Budziosz J, Stanek A, Sieroń A, Witkoś J, Cholewka A, Sieroń K. Effects of low-frequency electromagnetic field on oxidative stress in selected structures of the central nervous system. *Oxid Med Cell Longev.* 2018:1427412, 2018.

(E) (VT, AE, DOD, IAO, IX) Bułdak RJ, Polaniak R, Bułdak L, Zwirska-Korczala K, Skonieczna M, Monsiol A, Kukła M, Duława-Bułdak A, Birkner E. Short-term exposure to 50 Hz ELF-EMF alters the cisplatin-induced oxidative response in AT478 murine squamous cell carcinoma cells. *Bioelectromagnetics.* 33(8):641-651, 2012.

(E)(VT, AE, IFR) Calabrò E, Condello S, Currò M, Ferlazzo N, Caccamo D, Magazù S, Ientile R. Effects of low intensity static magnetic field on FTIR spectra and ROS production in SH-SY5Y neuronal-like cells. *Bioelectromagnetics.* 34(8):618-629, 2013.

(E) (VT, AE, IFR, IOD, DAO) Calcabrini C, Mancini U, De Bellis R, Diaz AR, Martinelli M, Cucchiarini L, Sestili P, Stocchi V, Potenza L. Effect of extremely low-frequency electromagnetic fields on antioxidant activity in the human keratinocyte cell line NCTC 2544. *Biotechnol Appl Biochem.* 64(3):415-422, 2017.

(E) (VT, AE, DFR) Calota V, Dragoiu S, Meghea A, Giurginca M. Decrease of luminol chemiluminescence upon exposure of human blood serum to 50 Hz electric fields. *Bioelectrochemistry.* 69(1):126-127, 2006.

(E)(VT, AE, IFR) Calota V, Dragoiu S, Meghea A, Giurginca M. Effects of prooxidants on human serum exposed to 50 Hz magnetic fields. *Electromagn Biol Med.*26(2):135-140, 2007.

(E)(VO, CE, DOD, IOD, DFR, IFR) Canseven AG, Coskun S, Seyhan N. Effects of various extremely low frequency magnetic fields on the free radical processes, natural antioxidant system and respiratory burst system activities in the heart and liver tissues. *Indian J Biochem Biophys.* 45(5):326-331, 2008.

(E) (VT, AE, IFR) Chen Y, Hong L, Zeng Y, Shen Y, Zeng Q. Power frequency magnetic fields induced reactive oxygen species-related autophagy in mouse embryonic fibroblasts. *Int J Biochem Cell Biol.* 57:108-114, 2014.

(E)(VT, AE, IX) Cheun BS, Yi SH, Baik KY, Lim JK, Yoo JS, Shin HW, Soh KS. Biophoton emission of MDCK cell with hydrogen peroxide and 60 Hz AC magnetic field. *J Environ Biol.* 28(4):735-740, 2007.

(E) (VT, AE, IFR, IOD, IAO) Chu LY, Lee JH, Nam YS, Lee YJ, Park WH, Lee BC, Kim D, Chung YH, Jeong JH. Extremely low frequency magnetic field induces oxidative stress in mouse cerebellum. *Gen Physiol Biophys.* 30(4):415-421, 2011.

(E) (VO, AE, IFR) Chung YH, Lee YJ, Lee HS, Chung SJ, Lim CH, Oh KW, Sohn UD, Park ES, Jeong JH. Extremely low frequency magnetic field modulates the level of neurotransmitters. *Korean J Physiol Pharmacol.* 19(1):15-20, 2015.

(E) (VO, HU, CE, IAO) Cichoń N, Bijak M, Miller E, Saluk J. Extremely low frequency electromagnetic field (ELF-EMF) reduces oxidative stress and improves functional and psychological status in ischemic stroke patients. *Bioelectromagnetics.* 38(5):386-396, 2017a.

(E) (HU, CE, IFR) Cichoń N, Czarny P, Bijak M, Miller E, Śliwiński T, Szemraj J, Saluk-Bijak J. Benign effect of extremely low-frequency electromagnetic field on brain plasticity assessed by nitric oxide metabolism during poststroke rehabilitation. *Oxid Med Cell Longev.* 2017:2181942, 2017b.

(E) (HU, CE, IAO) Cichon N, Bijak M, Synowiec E, Miller E, Sliwinski T, Saluk-Bijak J. Modulation of antioxidant enzyme gene expression by extremely low frequency electromagnetic field in post-stroke patients. *Scand J Clin Lab Invest.* 78(7-8):626-631, 2018a.

(E) (HU, CE, DOD) Cichoń N, Rzeźnicka P, Bijak M, Miller E, Miller S, Saluk J. Extremely low frequency electromagnetic field reduces oxidative stress during the rehabilitation of post-acute stroke patients. *Adv Clin Exp Med*. 27(9):1285-1293, 2018b.

(E) (VO, CE, IAO, DAO) Ciejka EB, Goraca A. The influence of low-frequency magnetic field on plasma antioxidant capacity and heart rate. *Wiad Lek*. 62(2):81-86, 2009.

(E) (VO, CE, IFR) Ciejka E, Skibska B, Kleniewska P, Goraca A. [Influence of low frequency magnetic field on chosen parameters of oxidative stress in rat's muscles]. *Pol Merkur Lekarski*. 29(174):361-364, 2010. [Article in Polish]

(E) (VO, CE, IOD) Ciejka E, Kleniewska P, Skibska B, Goraca A. Effects of extremely low frequency magnetic field on oxidative balance in brain of rats. *J Physiol Pharmacol*. 62(6):657-661, 2011.

(E) (VO, CE, IAO) Ciejka E, Jakubowska E, Zelechowska P, Huk-Kolega H, Kowalczyk A, Goraca A. [Effect of extremely low frequency magnetic field on glutathione in rat muscles]. *Med Pr*. 65(3):343-349, 2014. [Article in Polish]

(E) (VO, CE, DFR, DOD, IAO, IX) Coballase-Urrutia E, Navarro L, Ortiz JL, Verdugo-Díaz L, Gallardo JM, Hernández ME, Estrada-Rojo F. Static magnetic fields modulate the response of different oxidative stress markers in a restraint stress model animal. *Biomed Res Int*. 2018:3960408, 2018.

Consales C, Merla C, Marino C, Benassi B. Electromagnetic fields, oxidative stress, and neurodegeneration. *Int J Cell Biol*. 2012:683897, 2012. (Review)

(E) (VT, AE, IFR) Consales C, Cirotti C, Filomeni G, Panatta M, Butera A, Merla C, Lopresto V, Pinto R, Marino C, Benassi B. Fifty-hertz magnetic field affects the epigenetic modulation of the miR-34b/c in neuronal cells. *Mol Neurobiol*. 55(7):5698-5718, 2018.

- (NE) (IV, AE) Consales C, Panatta M, Butera A, Filomeni G, Merla C, Carrì MT, Marino C, Benassi B. 50-Hz magnetic field impairs the expression of iron-related genes in the in vitro SOD1^{G93A} model of amyotrophic lateral sclerosis. *Int J Radiat Biol.* 95:368-377, 2019.
- (E) (VO, CE, IFR, IOD) Coşkun S, Balabanli B, Canseven A, Seyhan N. Effects of continuous and intermittent magnetic fields on oxidative parameters in vivo. *Neurochem Res.* 34(2):238-243, 2009.
- (E) (VO, CE, IOD, DAO) Cui Y, Ge Z, Rizak JD, Zhai C, Zhou Z, Gong S, Che Y. Deficits in water maze performance and oxidative stress in the hippocampus and striatum induced by extremely low frequency magnetic field exposure. *PLoS One.* 7(5):e32196, 2012.
- (NE) (VT, AE) de Groot MW, Kock MD, Westerink RH. Assessment of the neurotoxic potential of exposure to 50 Hz extremely low frequency electromagnetic fields (ELF-EMF) in naïve and chemically-stressed PC12 cells. *Neurotoxicology.* 44:358-364, 2014.
- (NE) (VT, AE, IX) De Mattei M, Pasello M, Pellati A, Stabellini G, Massari L, Gemmati D, Caruso A. Effects of electromagnetic fields on proteoglycan metabolism of bovine articular cartilage explants. *Connect Tissue Res.* 44(3-4):154-159, 2003.
- (E)(VT, AE, IFR, IAO) De Nicola M, Cordisco S, Cerella C, Albertini MC, D'Alessio M, Accorsi A, Bergamaschi A, Magrini A, Ghibelli L. Magnetic fields protect from apoptosis via redox alteration. *Ann N Y Acad Sci.* 1090:59-68, 2006.
- (E) (VT, VO, AE, IOD, DAO, IX) Deng B, Xu H, Zhang J, Wang J, Han LC, Li LY, Wu GL, Hou YN, Guo GZ, Wang Q, Sang HF, Xu LX. Neuroprotective effects of sevoflurane against electromagnetic pulse-induced brain injury through inhibition of neuronal oxidative stress and apoptosis. *PLoS One.* 9(3):e91019, 2014.
- (E) (VO, CE, IOD, DAO) Deng Y, Zhang Y, Jia S, Liu J, Liu Y, Xu W, Liu L. Effects of aluminum and extremely low frequency electromagnetic radiation on oxidative stress and memory in brain of mice. *Biol Trace Elem Res.* 156(1-3):243-252, 2013.

(E) (VT, AE, DRF) Di S, Tian Z, Qian A, Li J, Wu J, Wang Z, Zhang D, Yin D, Brandi ML, Shang P. Large gradient high magnetic field affects FLG29.1 cells differentiation to form osteoclast-like cells. *Int J Radiat Biol.* 88(11):806-813, 2012.

(NE) (VT, CE) Di Loreto S, Falone S, Caracciolo V, Sebastiani P, D'Alessandro A, Mirabilio A, Zimmitti V, Amicarelli F. Fifty hertz extremely low-frequency magnetic field exposure elicits redox and trophic response in rat-cortical neurons. *J Cell Physiol.* 219(2):334-343, 2009.

(E) (VO, CE, IOD) Dinčić M, Krstić DZ, Čolović MB, Nešović Ostojić J, Kovačević S, De Luka SR, Djordjević DM, Ćirković S, Brkić P, Todorović J. Modulation of rat synaptosomal ATPases and acetylcholinesterase activities induced by chronic exposure to the static magnetic field. *Int J Radiat Biol.* 94(11):1062-1071, 2018

(E) (VT, AE, IX) Ding GR, Nakahara T, Hirose H, Koyama S, Takashima Y, Miyakoshi J. Extremely low frequency magnetic fields and the promotion of H₂O₂-induced cell death in HL-60 cells. *Int J Radiat Biol.* 80(4):317-324, 2004.

(E) (VO, CE, IFR) Djordjevic NZ, Paunović MG, Peulić AS. Anxiety-like behavioural effects of extremely low-frequency electromagnetic field in rats. *Environ Sci Pollut Res Int.* 24(27):21693-21699, 2017.

Dodson CA, Hore PJ, Wallace MI. A radical sense of direction: signalling and mechanism in cryptochrome magnetoreception. *Trends Biochem Sci.* 38(9):435-446, 2013. (Review)

(E) (IV, AE, IFR, DFR, IAO, DAO, IOD) Dornelles EB, Goncalves BD, Schott KL, Barbisan F, Unfer TC, Glanzner WG, Machado AK, Cadona FC, Azzolin VF, Montano MA, Griner J, da Cruz IB. Cytotoxic effects of moderate static magnetic field exposure on human periphery blood mononuclear cells are influenced by Val16Ala-MnSOD gene polymorphism. *Environ Sci Pollut Res Int.* 24(5):5078-5088, 2017.

(E) (VO, CE, IFR, DAO, IX) Duan Y, Wang Z, Zhang H, He Y, Lu R, Zhang R, Sun G, Sun X. The preventive effect of lotus seedpod procyanidins on cognitive impairment and oxidative damage induced by extremely low frequency electromagnetic field exposure. *Food Funct.* 4(8):1252-1262, 2013.

(NE) (VT, AE) Duan W, Liu C, Zhang L, He M, Xu S, Chen C, Pi H, Gao P, Zhang Y, Zhong M, Yu Z, Zhou Z. Comparison of the genotoxic effects induced by 50 Hz extremely low-frequency electromagnetic fields and 1800 MHz radiofrequency electromagnetic fields in GC-2 cells. *Radiat Res.* 183(3):305-314, 2015.

(E) (VT, AE, IX, DFR) Duong CN, Kim JY. Exposure to electromagnetic field attenuates oxygen-glucose deprivation-induced microglial cell death by reducing intracellular Ca²⁺ and ROS. *Int J Radiat Biol.* 92(4):195-201, 2016.

(E) (IV, AE, CE, IFR, DFR, IAO) Ehnert S, Fentz AK, Schreiner A, Birk J, Wilbrand B, Ziegler P, Reumann MK, Wang H, Falldorf K, Nussler AK. Pulsed electromagnetic fields cause antioxidative defense mechanisms in human osteoblasts via induction of •O₂⁻ and H₂O₂. *Sci Rep.* 7(1):14544, 2017.

(E) (VO, CE, IOD, IAO) Emre M, Cetiner S, Zencir S, Unlukurt I, Kahraman I, Topcu Z. Oxidative stress and apoptosis in relation to exposure to magnetic field. *Cell Biochem Biophys.* 59(2):71-77, 2011.

(E) (VO, CE, IFR) Erdal N, Gürgül S, Tamer L, Ayaz L. Effects of long-term exposure of extremely low frequency magnetic field on oxidative/nitrosative stress in rat liver. *J Radiat Res (Tokyo).* 49(2):181-187, 2008.

(E) (VT, AE, IAO, IX) Errico Provenzano A, Amatori S, Nasoni MG, Persico G, Russo S, Mastrogiacomo AR, Gambarara A, Fanelli M. Effects of fifty-hertz electromagnetic fields on granulocytic differentiation of ATRA-treated acute promyelocytic leukemia NB4 cells. *Cell Physiol Biochem.* 46(1):389-400, 2018.

(E) (IV, AE, IAO, IX, MC) Falone S, Grossi MR, Cinque B, D'Angelo B, Tettamanti E, Cimini A, Di Ilio C, Amicarelli F. Fifty hertz extremely low-frequency electromagnetic field causes changes in redox and differentiative status in neuroblastoma cells. *Int J Biochem Cell Biol.* 39(11):2093-2106, 2007.

(E) (VO, CE, IAO, DAO) Falone S, Mirabilio A, Carbone MC, Zimmiti V, Di Loreto S, Mariggì MA, Mancinelli R, Di Ilio C, Amicarelli F. Chronic exposure to 50 Hz magnetic fields causes a significant weakening of antioxidant defence systems in aged rat brain. *Int J Biochem Cell Biol.* 40(12):2762-2770, 2008.

(E) (VT, AE, IAO, IX) Falone S, Marchesi N, Osera C, Fassina L, Comincini S, Amadio M, Pascale A. Pulsed electromagnetic field (PEMF) prevents pro-oxidant effects of H₂O₂ in SK-N-BE(2) human neuroblastoma cells. *Int J Radiat Biol.* 92(5):281-286, 2016.

(E) (VT, AE, IAO) Falone S, Santini S Jr, Cordone V, Cesare P, Bonfigli A, Grannonico M, Di Emidio G, Tatone C, Cacchio M, Amicarelli F. Power frequency magnetic field promotes a more malignant phenotype in neuroblastoma cells via redox-related mechanisms. *Sci Rep.* 7(1):11470, 2017.

Falone S, Santini S Jr, Cordone V, Di Emidio G, Tatone C, Cacchio M, Amicarelli F. Extremely low-frequency magnetic fields and redox-responsive pathways linked to cancer drug resistance: Insights from co-exposure-based *In Vitro* studies. *Front Public Health.* 6:33, 2018. (Review)

(VT, AE, IFR, AO) Feng B, Qiu L, Ye C, Chen L, Fu Y, Sun W. Exposure to a 50-Hz magnetic field induced mitochondrial permeability transition through the ROS/GSK-3 β signaling pathway. *Int J Radiat Biol.* 92:148-155, 2016a.

(E) (VT, AE, IFR) Feng B, Dai A, Chen L, Qiu L, Fu Y, Sun W. NADPH oxidase-produced superoxide mediated a 50-Hz magnetic field-induced epidermal growth factor receptor clustering. *Int J Radiat Biol.* 92:596-602, 2016b.

(E) (VT, AE, IFR, MC) Feng B, Ye C, Qiu L, Chen L, Fu Y, Sun W. Mitochondrial ROS release and subsequent Akt Activation potentially mediated the anti-apoptotic effect of a 50-Hz magnetic field on FL cells. *Cell Physiol Biochem.* 38(6):2489-2499, 2016c.

(E) (VO, CE, DAO) Fernie KJ, Bird DM. Evidence of oxidative stress in American kestrels exposed to electromagnetic fields. *Environ Res.* 86(2):198-207, 2001.

(E) (VT, AE, IX) Fiorani M, Biagiarelli B, Vetrano F, Guidi G, Dachà M, Stocchi V. In vitro effects of 50 Hz magnetic fields on oxidatively damaged rabbit red blood cells. *Bioelectromagnetics.* 18(2):125-131, 1997.

(E) (VT, AE, IFR) Fitzsimmons RJ, Gordon SL, Kronberg J, Ganey T, Pilla AA. A pulsing electric field (PEF) increases human chondrocyte proliferation through a transduction pathway involving nitric oxide signaling. *J Orthop Res.* 26(6):854-859, 2008.

- (E) (VT, AE, IFR) Frahm J, Lantow M, Lupke M, Weiss DG, Simkó M. Alteration in cellular functions in mouse macrophages after exposure to 50 Hz magnetic fields. *J Cell Biochem.* 99(1):168-177, 2006.
- (E) (VT, AE, IFR) Frahm J, Mattsson MO, Simkó M. Exposure to ELF magnetic fields modulate redox related protein expression in mouse macrophages. *Toxicol Lett.* 192(3):330-336, 2010.
- (E) (VT, AE, IFR) Garip AI, Akan Z. Effect of ELF-EMF on number of apoptotic cells; correlation with reactive oxygen species and HSP. *Acta Biol Hung.* 61(2):158-167, 2010.
- (E)(VO, CE, IAO, IX) Ghodbane S, Amara S, Arnaud J, Garrel C, Faure H, Favier A, Sakly M, Abdelmelek H. Effect of selenium pre-treatment on plasma antioxidant vitamins A (retinol) and E (α -tocopherol) in static magnetic field-exposed rats. *Toxicol Ind Health.* 27(10):949-955, 2011a.
- (E) (VO, CE, IAO, DAO) Ghodbane S, Amara S, Garrel C, Arnaud J, Ducros V, Favier A, Sakly M, Abdelmelek H. Selenium supplementation ameliorates static magnetic field-induced disorders in antioxidant status in rat tissues. *Environ Toxicol Pharmacol.* 31(1):100-106, 2011b.
- Ghodbane S, Lahbib A, Sakly M, Abdelmelek H. Bioeffects of static magnetic fields: oxidative stress, genotoxic effects, and cancer studies. *Biomed Res Int.* 2013:602987, 2013. (Review)
- (E) (VO, CE, IX) Ghodbane S1, Amara S, Lahbib A, Louchami K, Sener A, Sakly M, Abdelmelek H. Vitamin E prevents glucose metabolism alterations induced by static magnetic field in rats. *Environ Sci Pollut Res Int.* 21(22):12731-12738, 2014.
- (E) (VO, CE, IAO, AO) Ghodbane S, Ammari M, Lahbib A, Sakly M, Abdelmelek H. Static magnetic field exposure-induced oxidative response and caspase-independent apoptosis in rat liver: effect of selenium and vitamin E supplementations. *Environ Sci Pollut Res Int.* 22(20):16060-16066, 2015a.
- (E) (VO, CE, IAO, IOD, AO) Ghodbane S, Lahbib A, Ammari M, Sakly M, Abdelmelek H. Does static magnetic field-exposure induced oxidative stress and apoptosis in rat kidney and muscle? Effect of vitamin E and selenium supplementations. *Gen Physiol Biophys.* 34(1):23-32, 2015b.

(NE)(VT, AE, IX) Giorgi G, Lecciso M, Capri M, Lukas Yani S, Virelli A, Bersani F, Del Re B. An evaluation of genotoxicity in human neuronal-type cells subjected to oxidative stress under an extremely low frequency pulsed magnetic field. *Mutat Res Genet Toxicol Environ Mutagen.* 775-776:31-37, 2014.

(E) (VT, AE, IOD, IX) Giorgi G, Pirazzini C, Bacalini MG, Giuliani C, Garagnani P, Capri M, Bersani F, Del Re B. Assessing the combined effect of extremely low-frequency magnetic field exposure and oxidative stress on LINE-1 promoter methylation in human neural cells. *Radiat Environ Biophys.* 56(2):193-200, 2017.

(E) (VO, CE, DOD, IAO) Glinka M, Sieroń A, Birkner E, Cieślar G. Influence of extremely low-frequency magnetic field on the activity of antioxidant enzymes during skin wound healing in rats. *Electromagn Biol Med.* 32(4):463-470, 2013.

(E) (IV, AE, DAO) Glinka M, Gawron S, Sieroń A, Pawłowska-Góral K, Cieślar G, Sieroń K. Impact of static magnetic field on the antioxidant defence system of mice fibroblasts. *Biomed Res Int.* 2018:5053608, 2018.

Ghodbane S, Lahbib A, Sakly M, Abdelmelek H. Bioeffects of static magnetic fields: oxidative stress, genotoxic effects, and cancer studies. *Biomed Res Int.* 2013:602987, 2013. (Review)

(E) (VO, CE, IOD) Gok DK, Akpınar D, Hidisoglu E, Ozen S, Agar A, Yargicoglu P. The developmental effects of extremely low frequency electric fields on visual and somatosensory evoked potentials in adult rats. *Electromagn Biol Med.* 35(3):245-259, 2016.

(E) (VT, CE, IRF, IOD, DAO) Goraca A, Ciejka E, Piechota A. Effects of extremely low frequency magnetic field on the parameters of oxidative stress in heart. *J Physiol Pharmacol.* 61(3):333-338, 2010.

(E) (VO, CE, IFR, IOD, DAO) Guler G, Turkozer Z, Tomruk A, Seyhan N. The protective effects of N-acetyl-L-cysteine and epigallocatechin-3-gallate on electric field-induced hepatic oxidative stress. *Int J Radiat Biol.* 84(8):669-680, 2008.

(NE) (VO, CE) Güler G, Türközer Z, Ozgur E, Tomruk A, Seyhan N, Karasu C. Protein oxidation under extremely low frequency electric field in guinea pigs. Effect of N-acetyl-L-cysteine treatment. *Gen Physiol Biophys.* 28(1):47-55, 2009a.

- (NE) (VO, CE) Güler G, Türközer Z, Ozgur E, Seyhan N. Antioxidants alleviate electric field-induced effects on lung tissue based on assays of heme oxygenase-1, protein carbonyl content, malondialdehyde, nitric oxide, and hydroxyproline. *Sci Total Environ.* 407(4):1326-1332, 2009b.
- (E) (VT, AE, IAO, IFC, IX) Hajipour Verdom B, Abdolmaleki P, Behmanesh M. The static magnetic field remotely boosts the efficiency of doxorubicin through modulating ROS behaviors. *Sci Rep.* 8(1):990, 2018.
- (E) (VO, CE, IAO, LI) Hajnorouzi A, Vaezzadeh M, Ghanati F, Jamnezhad H, Nahidian B. Growth promotion and a decrease of oxidative stress in maize seedlings by a combination of geomagnetic and weak electromagnetic fields. *J Plant Physiol.* 168(10):1123-1128, 2011.
- (E) (VO, AE, IOD, IAO) Hanini R, Chatti A, Ghorbel SB, Landoulsi A. Role of SOD gene in response to static magnetic fields in *Pseudomonas aeruginosa*. *Curr Microbiol.* 74(8):930-937, 2017.
- (NE) (VO, CE) Harakawa S, Inoue N, Hori T, Tochio K, Kariya T, Takahashi K, Doge F, Suzuki H, Nagasawa H. Effects of a 50 Hz electric field on plasma lipid peroxide level and antioxidant activity in rats. *Bioelectromagnetics.* 26(7):589-594, 2005.
- (E) (VO, CE, IOD, DAO) Hashish AH, El-Missiry MA, Abdelkader HI, Abou-Saleh RH. Assessment of biological changes of continuous whole body exposure to static magnetic field and extremely low frequency electromagnetic fields in mice. *Ecotoxicol Environ Saf.* 71(3):895-902. 2008.
- (E) (VT, AE, IOD, IFR, IAO, DAO) Henrykowska G, Jankowski W, Pacholski K, Lewicka M, Smigielski J, Dziedziczak-Buczyńska M, Buczyński A. The effect of 50 Hz magnetic field of different shape on oxygen metabolism in blood platelets: in vitro studies. *Int J Occup Med Environ Health.* 22(3):269-276, 2009.
- (NE) (VT, AE) Hong MN, Han NK, Lee HC, Ko YK, Chi SG, Lee YS, Gimm YM, Myung SH, Lee JS. Extremely low frequency magnetic fields do not elicit oxidative stress in MCF10A cells. *Radiat Res.* 53(1):79-86, 2012.
- (E) (HU, CE, IOD, IAO) Hosseinabadi MB, Khanjani N. The effect of extremely low-frequency electromagnetic fields on the prevalence of musculoskeletal disorders and the role of oxidative stress. *Bioelectromagnetics.* 2019 Jun 18. doi: 10.1002/bem.22198. [Epub ahead of print]

- (E) (VT, AE, IFR) Höytö A, Herrala M, Luukkonen J, Juutilainen J, Naarala J. Cellular detection of 50 Hz magnetic fields and weak blue light: effects on superoxide levels and genotoxicity. *Int J Radiat Biol.* 7:1-7, 2017.
- (E) (VO, CE, DFR) Hu Y, Lai J, Wan B, Liu X, Zhang Y, Zhang J, Sun D, Ruan G, Liu E, Liu GP, Chen C, Wang DW. Long-term exposure to ELF-MF ameliorates cognitive deficits and attenuates tau hyperphosphorylation in 3xTg AD mice. *Neurotoxicology* 53:290-300, 2016.
- (E) (VT, AE, IX, AO) Jajte J, Zmysłony M, Palus J, Dziubałtowska E, Rajkowska E. Protective effect of melatonin against in vitro iron ions and 7 mT 50 Hz magnetic field-induced DNA damage in rat lymphocytes. *Mutat Res.* 483(1-2):57-64, 2001.
- (E) (VT, AE, IX) Jajte J, Grzegorzczak J, Zmysłony M, Rajkowska E. Effect of 7 mT static magnetic field and iron ions on rat lymphocytes: apoptosis, necrosis and free radical processes. *Bioelectrochemistry.* 57(2):107-111, 2002.
- (E) (VT, AE, IX, AO) Jajte J, Zmysłony M, Rajkowska E. [Protective effect of melatonin and vitamin E against prooxidative action of iron ions and static magnetic field]. *Med Pr.* 54(1):23-28, 2003. [Article in Polish]
- (E) (VO, CE, IFR, IOD, IAO) Jelenković A, Janać B, Pesić V, Jovanović DM, Vasiljević I, Prolić Z. Effects of extremely low-frequency magnetic field in the brain of rats. *Brain Res Bull.* 68(5):355-360, 2006.
- (E) (VO, AE, IFR) Jeong JH, Kum C, Choi HJ, Park ES, Sohn UD. Extremely low frequency magnetic field induces hyperalgesia in mice modulated by nitric oxide synthesis. *Life Sci.* 78(13):1407-1412, 2006.
- (NE) (VT, AE) Jin H, Yoon HE, Lee JS, Kim JK, Myung SH, Lee YS. Effects on g2/m phase cell cycle distribution and aneuploidy formation of exposure to a 60 Hz electromagnetic field in combination with ionizing radiation or hydrogen peroxide in I132 nontumorigenic human lung epithelial cells. *Korean J Physiol Pharmacol.* 19(2):119-124, 2015.
- (NE) (VT, AE) Jin YB, Kang GY, Lee JS, Choi JI, Lee JW, Hong SC, Myung SH, Lee YS. Effects on micronuclei formation of 60-Hz electromagnetic field exposure with ionizing radiation, hydrogen peroxide, or c-Myc overexpression. *Int J Radiat Biol.* 88(4):374-80, 2012.

(NE) (VT, AE) Jin YB, Choi SH, Lee JS, Kim JK, Lee JW, Hong SC, Myung SH, Lee YS. Absence of DNA damage after 60-Hz electromagnetic field exposure combined with ionizing radiation, hydrogen peroxide, or c-Myc overexpression. *Radiat Environ Biophys*. 53(1):93-101, 2014.

(E) (VO, CE, IOD, DAO) Jouni FJ, Abdolmaleki P, Ghanati F. Oxidative stress in broad bean (*Vicia faba* L.) induced by static magnetic field under natural radioactivity. *Mutat Res*. 741(1-2):116-121, 2012.

(E) (VT, AE, IFR, IX) Kamalipooya S, Abdolmaleki P, Salemi Z, Javani Jouni F, Zafari J, Soleimani H. Simultaneous application of cisplatin and static magnetic field enhances oxidative stress in HeLa cell line. *In Vitro Cell Dev Biol Anim*. 53(9):783-790, 2017.

(E) (VO, CE, IOD) Kantar Gok D, Akpınar D, Yargıoğlu P, Ozen S, Aslan M, Demir N, Derin N, Agar A. Effects of extremely low-frequency electric fields at different intensities and exposure durations on mismatch negativity. *Neuroscience*. 272C:154-166, 2014.

(E) (VO, CE, IFR, IAO, IOD) Karimi SA, Salehi I, Shykhi T, Zare S, Komaki A. Effects of exposure to extremely low-frequency electromagnetic fields on spatial and passive avoidance learning and memory, anxiety-like behavior and oxidative stress in male rats. *Behav Brain Res*. 359:630-638, 2019.

(E) (VO, AE, IFR) Kavaliers M, Choleris E, Prato FS, Ossenkopp K. Evidence for the involvement of nitric oxide and nitric oxide synthase in the modulation of opioid-induced antinociception and the inhibitory effects of exposure to 60-Hz magnetic fields in the land snail. *Brain Res*. 809(1):50-57, 1998.

(NE) (VT, AE) Kesari KK, Luukkonen J, Juutilainen J, Naarala J. Genomic instability induced by 50 Hz magnetic fields is a dynamically evolving process not blocked by antioxidant treatment. *Mutat Res Genet Toxicol Environ Mutagen*. 794:46-51, 2015.

(E) (VT, AE, IFR) Kesari KK, Juutilainen J, Luukkonen J, Naarala J. Induction of micronuclei and superoxide production in neuroblastoma and glioma cell lines exposed to weak 50 Hz magnetic fields. *J R Soc Interface*. 2016 Jan; 13(114). pii: 20150995. doi: 10.1098/rsif.2015.0995.

(E) (VT, AE, IFR) Khadir R, Morgan JL, Murray JJ. Effects of 60 Hz magnetic field exposure on polymorphonuclear leukocyte activation. *Biochim Biophys Acta*. 1472(1-2):359-367, 1999.

(E) (VT, AE, IFR, AO) Kim SJ, Jang YW, Hyung KE, Lee DK, Hyun KH, Jeong SH, Min KH, Kang W, Jeong JH, Park SY, Hwang KW. Extremely low-frequency electromagnetic field exposure enhances inflammatory response and inhibits effect of antioxidant in RAW 264.7 cells. *Bioelectromagnetics*. 38(5):374-385, 2017.

(E) (VT,AE, IX, MC) Kimsa-Dudek M, Synowiec-Wojtarowicz A, Derewniuk M, Gawron S, Paul-Samojedny M, Kruszniewska-Rajs C, Pawłowska-Góral K. Impact of fluoride and a static magnetic field on the gene expression that is associated with the antioxidant defense system of human fibroblasts. *Chem Biol Interact*. 287:13-19, 2018.

Kivrak EG, Yurt KK, Kaplan AA, Alkan I, Altun G. Effects of electromagnetic fields exposure on the antioxidant defense system. *J Microsc Ultrastruct*. 5(4):167-176, 2017. (Review)

(E) (VT, AE, IFR, AO) Koh EK, Ryu BK, Jeong DY, Bang IS, Nam MH, Chae KS. A 60-Hz sinusoidal magnetic field induces apoptosis of prostate cancer cells through reactive oxygen species. *Int J Radiat Biol*. 84(11):945-955, 2008.

(E) (VT, AE, IX) Koyama S, Nakahara T, Hirose H, Ding GR, Takashima Y, Isozumi Y, Miyakoshi J. ELF electromagnetic fields increase hydrogen peroxide (H₂O₂)-induced mutations in pTN89 plasmids. *Mutat Res*. 560(1):27-32, 2004.

(E) (VT, AE, IX) Koyama S, Sakurai T, Nakahara T, Miyakoshi J. Extremely low frequency (ELF) magnetic fields enhance chemically induced formation of apurinic/apyrimidinic (AP)sites in A172 cells. *Int J Radiat Biol*. 84(1):53-59, 2008.

(E) (VO, AE, IAO, DAO, IOD) Kthiri A, Hidouri S, Wiem T, Jeridi R, Sheehan D, Landouls A. Biochemical and biomolecular effects induced by a static magnetic field in *Saccharomyces cerevisiae*: Evidence for oxidative stress. *PLoS One*. 14(1):e0209843, 2019.

(E) (HU, CE, IFR) Kunt H, Şentürk İ, Gönül Y, Korkmaz M, Ahsen A, Hazman Ö, Bal A, Genç A, Songur A. Effects of electromagnetic radiation exposure on bone mineral density, thyroid, and oxidative stress index in electrical workers. *OncoTargets and Therapy*. 2016(9):745-754, 2016.

- (E) (VT, CE, IX) Kurzeja E, Synowiec-Wojtarowicz A, Stec M, Glinka M, Gawron S, Pawłowska-Góral K. Effect of a static magnetic fields and fluoride ions on the antioxidant defense system of mice fibroblasts. *Int J Mol Sci.*14(7):15017-15028, 2013.
- (E) (VO, CE, IFR, IOD, DAO) Kuzay D, Ozer C, Sirav B, Canseven AG, Seyhan N. Oxidative effects of extremely low frequency magnetic field and radio frequency radiation on testes tissues of diabetic and healthy rats. *Bratisl Lek Listy.* 118(5):278-282, 2017.
- Lahbib A, Ghodbane S, Sakly M, Abdelmelek H. Vitamins and glucose metabolism: The role of static magnetic fields. *Int J Radiat Biol.* 90(12):1240-1245, 2014. (Review)
- (E) (VO, AE, AO) Lai H, Singh NP. Melatonin and N-tert-butyl-alpha-phenylnitronone block 60-Hz magnetic field-induced DNA single and double strand breaks in rat brain cells. *J Pineal Res.* 22(3):152-162, 1997.
- (E) (VO, AE, AO) Lai H, Singh NP. Magnetic-field-induced DNA strand breaks in brain cells of the rat. *Environ Health Perspect.* 112(6):687-694, 2004.
- (E)(VT, AE, AO) Lai HC, Chan HW, Singh NP. Effects of radiation from a radiofrequency identification (RFID) microchip on human cancer cells. *Int J Radiat Biol.* 2016;92(3):156-161, 2016.
- (E) (VO, AE, IOD, IAO) Lee BC, Johng HM, Lim JK, Jeong JH, Baik KY, Nam TJ, Lee JH, Kim J, Sohn UD, Yoon G, Shin S, Soh KS. Effects of extremely low frequency magnetic field on the antioxidant defense system in mouse brain: a chemiluminescence study. *J Photochem Photobiol B.* 73(1-2):43-48, 2004.
- (NE) (VT, AE) Lee HJ, Jin YB, Lee JS, Choi JI, Lee JW, Myung SH, Lee YS. Combined effects of 60 Hz electromagnetic field exposure with various stress factors on cellular transformation in NIH3T3 cells. *Bioelectromagnetics.* 33(3):207-214, 2012.
- (E) (VT, AE, IRF) Lee HM, Kwon UH, Kim H, Kim HJ, Kim B, Park JO, Moon ES, Moon SH. Pulsed electromagnetic field stimulates cellular proliferation in human intervertebral disc cells. *Yonsei Med J.* 51(6):954-959, 2010.

Lewczuk B, Redlarski G, Zak A, Ziółkowska N, Przybylska-Gornowicz B, Krawczuk M. Influence of electric, magnetic, and electromagnetic fields on the circadian system: current stage of knowledge. *Biomed Res Int.* 2014;169459, 2014. (Review)

(E) (VT, AE, IOD, IAO) Lewicka M, Henrykowska GA, Pacholski K, Szczesny A, Dziedziczak-Buczyńska M, Buczyński A. The impact of electromagnetic radiation of different parameters on platelet oxygen metabolism - in vitro studies. *Adv Clin Exp Med.* 24(1):31-35, 2015.

(NE) (HU, CE) Li L, Xiong DF, Liu JW, Li ZX, Zeng GC, Li HL. A cross-sectional study on oxidative stress in workers exposed to extremely low frequency electromagnetic fields. *Int J Radiat Biol.* 91(5):420-425, 2015.

(E) (VO, AE, CE, IAO) Li SS, Zhang ZY, Yang CJ, Lian HY, Cai P. Gene expression and reproductive abilities of male *Drosophila melanogaster* subjected to ELF-EMF exposure. *Mutat Res.* 758(1-2):95-103, 2013.

(E) (IV AE, IFR, IAO) Lian HY, Lin KW, Yang C, Cai P. Generation and propagation of yeast prion [URE3] are elevated under electromagnetic field. *Cell Stress Chaperones.* 23(4):581-594, 2018.

(E) (VT, AE, AO) Liu DD, Ren Z, Yang G, Zhao QR, Mei YA. Melatonin protects rat cerebellar granule cells against electromagnetic field-induced increases in Na⁺ currents through intracellular Ca²⁺ release. *J Cell Mol Med.* 18(6):1060-1070, 2014.

(E) (VO, CE, IOD, DAO) Liu Y, Weng E, Zhang Y, Hong R. [Effects of extremely low frequency electromagnetic field and its combination with lead on the antioxidant system in mouse] *Zhonghua Lao Dong Wei Sheng Zhi Ye Bing Za Zhi.* 20(4):263-265, 2002. [Article in Chinese]

(E) (VO, CE, IOD, DAO, AO) Luo X, Chen M, Duan Y, Duan W, Zhang H, He Y, Yin C, Sun G, Sun X. Chemoprotective action of lotus seedpod procyanidins on oxidative stress in mice induced by extremely low-frequency electromagnetic field exposure. *Biomed Pharmacother.* 82:640-648, 2016.

(E) (VO, AE, DAO) Luo K, Luo C, Li G, Yao X, Gao R, Hu Z, Zhang G, Zhao H. High-voltage electrostatic field-induced oxidative stress: Characterization of the physiological effects in *Sitobion avenae* (Hemiptera: Aphididae) across multiple generations. *Bioelectromagnetics.* 40(1):52-61, 2019.

- (E) (VO, AC, IFR) Lupke M, Rollwitz J, Simkó M. Cell activating capacity of 50 Hz magnetic fields to release reactive oxygen intermediates in human umbilical cord blood-derived monocytes and in Mono Mac 6 cells. *Free Radic Res.* 38(9):985-993, 2004.
- (E) (VT, AE, IFR) Luukkonen J, Liimatainen A, Juutilainen J, Naarala J. Induction of genomic instability, oxidative processes, and mitochondrial activity by 50 Hz magnetic fields in human SH-SY5Y neuroblastoma cells. *Mutat Res.* 760:33-41, 2014.
- (E) (VT, AE, MC) Mahmoudinasab H, Sanie-Jahromi F, Saadat M. Effects of extremely low-frequency electromagnetic field on expression levels of some antioxidant genes in human MCF-7 cells. *Mol Biol Res Commun.* 5(2):77-85. 2016.
- (E) (VT, AE, IAO, IX) Mahmoudinasab H, Saadat M. Electromagnetic field could protect SH-SY5Y cells against cisplatin cytotoxicity, but not MCF-7 cells. *DNA Cell Biol.* 37(4):330-335, 2018a.
- (E) (VT, AE, IX, MC) Mahmoudinasab H, Saadat M. Expressions of some antioxidant genes in SH-SY5Y cells treated with β -lapachone, morphine and electromagnetic field. *Mol Biol Rep.* 45(3):379-387, 2018b.
- (E) (VO, CE, TOD, DAO) Maliszewska J, Marciniak P, Kletkiewicz H, Wyszowska J, Nowakowska A, Rogalska J. Electromagnetic field exposure (50 Hz) impairs response to noxious heat in American cockroach. *J Comp Physiol A Neuroethol Sens Neural Behav Physiol.* 2018 May 2. doi: 10.1007/s00359-018-1264-2. [Epub ahead of print]
- (E) (VO, CE, IFR, IOD) Manikonda PK, Rajendra P, Devendranath D, Gunasekaran B, Channakeshava, Aradhya SR, Sashidhar RB, Subramanyam C. Extremely low frequency magnetic fields induce oxidative stress in rat brain. *Gen Physiol Biophys.* 33(1):81-90, 2014.
- (E) (VT AE, IFR, AO) Mannerling AC, Simkó M, Mild KH, Mattsson MO. Effects of 50-Hz magnetic field exposure on superoxide radical anion formation and HSP70 induction in human K562 cells. *Radiat Environ Biophys.* 49(4):731-741, 2010.

(NE) (VT, AE) Markkanen A, Naarala J, Juutilainen J. A Study on the effects of 50 Hz magnetic fields on UV-induced radical reactions in murine fibroblasts. *J Radiat Res (Tokyo)*. 51(5):609-613, 2010.

(E) (VT, AE, AO) Martínez MA, Úbeda A, Moreno J, Trillo MÁ. Power frequency magnetic fields affect the p38 MAPK-mediated regulation of NB69 cell proliferation implication of free radicals. *Int J Mol Sci*. 17(4):510, 2016.

(E) (VO, AE, DAO) Martínez-Sámano J, Torres-Durán PV, Juárez-Oropeza MA, Elías-Viñas D, Verdugo-Díaz L. Effects of acute electromagnetic field exposure and movement restraint on antioxidant system in liver, heart, kidney and plasma of Wistar rats: a preliminary report. *Int J Radiat Biol*. 86(12):1088-1094, 2010.

(E) (VO, AE, DAO) Martínez-Sámano J, Torres-Durán PV, Juárez-Oropeza MA, Verdugo-Díaz L. Effect of acute extremely low frequency electromagnetic field exposure on the antioxidant status and lipid levels in rat brain. *Arch Med Res*. 43(3):183-189, 2012.

(E) (VO, CE, IOD) Martínez-Sámano J, Flores-Poblano A, Verdugo-Díaz L, Juárez-Oropeza MA, Torres-Durán PV. Extremely low frequency electromagnetic field exposure and restraint stress induce changes on the brain lipid profile of Wistar rats. *BMC Neurosci*. 19(1):31, 2018.

(E) (VT, AE, AO) Martino CF. Static magnetic field sensitivity of endothelial cells. *Bioelectromagnetics*. 32(6):506-508, 2011.

(E) (VT, AE, IFR, LI) Martino CF, Castello PR. Modulation of hydrogen peroxide production in cellular systems by low level magnetic fields. *PLoS One*. 6(8):e22753, 2011.

(E) (VO, CE, DOD, IAO, IX) Medina-Fernandez FJ, Escribano BM, Agüera E, Aguilar-Luque M, Feijoo M, Luque E, Garcia-Maceira FI, Pascual-Leone A, Drucker-Colin R, Tunes I. Effects of transcranial magnetic stimulation on oxidative stress in experimental autoimmune encephalomyelitis. *Free Radic Res*. 51(5):460-469, 2017.

(E) (VO, CE, DOD, IAO, IX) Medina-Fernandez FJ, Escribano BM, Luque E, Caballero-Villarraso J, Gomez-Chaparro JL, Feijoo M, Garcia-Maceira FI, Pascual-Leone A, Drucker-Colin R, Tunes I. Comparative of transcranial magnetic stimulation and other treatments in experimental autoimmune encephalomyelitis. *Brain Res Bull*. 137:140-145, 2018.

Medina-Fernández FJ, Escribano BM, Padilla-Del-Campo C, Drucker-Colín R, Pascual-Leone Á, Túnez I. Transcranial magnetic stimulation as an antioxidant. *Free Radic Res.* 52(4):381-389, 2018. (Review)

(E) (VT, AE, IFR, MC) Merla C, Liberti M, Consales C, Denzi A, Apollonio F, Marino C, Benassi B. Evidences of plasma membrane-mediated ROS generation upon ELF exposure in neuroblastoma cells supported by a computational multiscale approach. *Biochim Biophys Acta Biomembr.* 1861(8):1446-1457, 2019.

(NE) (VT, AE) Messiha HL, Wongnate T, Chaiyen P, Jones AR, Scrutton NS. Magnetic field effects as a result of the radical pair mechanism are unlikely in redox enzymes. *J R Soc Interface.* 2015 Feb 6; 12(103). pii: 20141155. doi: 10.1098/rsif.2014.1155.

(E) (VO, CE, DAO) Miao X, Wang Y, Lang H, Lin Y, Guo Q, Yang M, Guo J, Zhang Y, Zhang J, Liu J, Liu Y, Zeng L, Guo G. Preventing electromagnetic pulse irradiation damage on testis using selenium-rich *Cordyceps* fungi. A preclinical study in young male mice. *OMICS.* 21(2):81-89, 2017.

(E) (VT, AE, IFR, IAO) Miliša M, Đikić D, Mandić T, Grozić D, Čolić I, Ostojić A. Response of aquatic protists to electric field exposure. *Int J Radiat Biol.* 93(8):818-830, 2017.

(E) (VT, AE, IFR) Mohammadi F, Ghanati F, Sharifi M, Chashmi NA. On the mechanism of the cell cycle control of suspension-cultured tobacco cells after exposure to static magnetic field. *Plant Sci.* 277:139-144. 2018.

Montoya RD. Magnetic fields, radicals and cellular activity. *Electromagn Biol Med.* 36(1):102-113, 2017. (Review)

(E) (VT, AE, CE, IFR, IAO, DAO) Morabito C, Guarnieri S, Fanò G, Marigliò MA. Effects of acute and chronic low frequency electromagnetic field exposure on PC12 cells during neuronal differentiation. *Cell Physiol Biochem.* 26(6):947-958, 2010a.

(E) (VT, AE, IFR, IAO) Morabito C, Rovetta F, Bizzarri M, Mazzoleni G, Fanò G, Marigliò MA. Modulation of redox status and calcium handling by extremely low frequency electromagnetic fields in C2C12 muscle cells: A real-time, single-cell approach. *Free Radic Biol Med.* 48(4):579-589, 2010b.

(E) (VT, AE, IFR) Naarala J, Kesari KK, McClure I, Chavarriaga C, Juutilainen J, Martino CF. Direction-dependent effects of combined static and ELF magnetic fields on cell proliferation and superoxide radical production. *Biomed Res Int.* 2017:5675086, 2017.

(NE) (VT, AE) Nakayama M, Nakamura A, Hondou T, Miyata H. Evaluation of cell viability, DNA single-strand breaks, and nitric oxide production in LPS-stimulated macrophage RAW264 exposed to a 50-Hz magnetic field. *Int J Radiat Biol.* 92(10):583-589, 2016.

(E) (VT, AE, IFR) Noda Y, Mori A, Liburdy RP, Packer L. Pulsed magnetic fields enhance nitric oxide synthase activity in rat cerebellum. *Pathophysiology.* 7(2):127-130, 2000.

Okano H. Effects of static magnetic fields in biology: role of free radicals. *Front Biosci.* 13:6106-6125, 2008. (Review)

(E) (VT, AE, IAO) Osera C, Fassina L, Amadio M, Venturini L, Buoso E, Magenes G, Govoni S, Ricevuti G, Pascale A. Cytoprotective response induced by electromagnetic stimulation on SH-SY5Y human neuroblastoma cell line. *Tissue Eng Part A.* 17(19-20):2573-2582, 2011.

(E) (VT, AE, IAO, IX) Osera C, Amadio M, Falone S, Fassina L, Magenes G, Amicarelli F, Ricevuti G, Govoni S, Pascale A. Pre-exposure of neuroblastoma cell line to pulsed electromagnetic field prevents H₂O₂-induced ROS production by increasing MnSOD activity. *Bioelectromagnetics.* 36(3):219-232, 2015.

(E) (VT, AE, IFR) Pakhomova ON, Khorokhorina VA, Bowman AM, Rodaitė-Riševičienė R, Saulis G, Xiao S, Pakhomov AG. Oxidative effects of nanosecond pulsed electric field exposure in cells and cell-free media. *Arch Biochem Biophys.* 527(1):55-64, 2012.

(E) (VO, AE, IOD, DAO) Pandir D, Sahingoz R. Magnetic field-induced oxidative stress and DNA damage in Mediterranean flour moth *Ephesia kuehniella* Zeller (Lepidoptera: Pyralidae) larvae. *J Pest Sci* 87(1): 79-87, 2014.

- (E) (VT, AE, IFR) Park JE, Seo YK, Yoon HH, Kim CW, Park JK, Jeon S. Electromagnetic fields induce neural differentiation of human bone marrow derived mesenchymal stem cells via ROS mediated EGFR activation. *Neurochem Int.* 62(4):418-424, 2013.
- (E) (VT, AE, IFR, DFR, IAO, DAO) Patruno A, Amerio P, Pesce M, Vianale G, Di Luzio S, Tulli A, Franceschelli S, Grilli A, Muraro R, Reale M. Extremely low frequency electromagnetic fields modulate expression of inducible nitric oxide synthase, endothelial nitric oxide synthase and cyclooxygenase-2 in the human keratinocyte cell line HaCat: potential therapeutic effects in wound healing. *Br J Dermatol.* 162(2):258-266, 2010.
- (E) (VT, AE, IAO, IFR) Patruno A, Tabrez S, Amerio P, Pesce M, Vianale G, Franceschelli S, Grilli A, Kamal MA, Reale M. Kinetic study on the effects of extremely low frequency electromagnetic field on catalase, cytochrome P450 and inducible nitric oxide synthase in human HaCaT and THP-1 cell lines. *CNS Neurol Disord Drug Targets.* 10(8):936-944, 2011.
- (E) (VT, AE, IFR, DAO) Patruno A, Pesce M, Marrone A, Speranza L, Grilli A, De Lutiis MA, Felaco M, Reale M. Activity of matrix metallo proteinases (MMPs) and the tissue inhibitor of MMP (TIMP)-1 in electromagnetic field-exposed THP-1 cells. *J Cell Physiol.* 227(6):2767-2774, 2012.
- (E) (VT, AE, IAO) Patruno A, Tabrez S, Pesce M, Shakil S, Kamal MA, Reale M. Effects of extremely low frequency electromagnetic field (ELF-EMF) on catalase, cytochrome P450 and nitric oxide synthase in erythro-leukemic cells. *Life Sci.* 121:117-123, 2015.
- (E) (IV, AE, IFR) Pilla AA. Electromagnetic fields instantaneously modulate nitric oxide signaling in challenged biological systems. *Biochem Biophys Res Commun.* 426(3):330-333, 2012.
- (E) (VO, CE, IAO, IX) Politański P, Rajkowska E, Pawlaczyk-Łuszczczyńska M, Dudarewicz A, Wiktorek-Smagur A, Sliwińska-Kowalska M, Zmysłony M. Static magnetic field affects oxidative stress in mouse cochlea. *Int J Occup Med Environ Health.* 23(4):377-384, 2010.
- (E) (VT, AE, DFR, IFR) Poniedziałek B, Rzymiski P, Nawrocka-Bogusz H, Jaroszyk F, Wiktorowicz K. The effect of electromagnetic field on reactive oxygen species production in human neutrophils in vitro. *Electromagn Biol Med.* 32(3):333-341, 2013a.

(E) (VT, AE, DFR, IFR) Poniedziałek B, Rzymiski P, Karczewski J, Jaroszyk F, Wiktorowicz K. Reactive oxygen species (ROS) production in human peripheral blood neutrophils exposed in vitro to static magnetic field. *Electromagn Biol Med.* 32(4):560-568, 2013b.

(E) (VT, AE, IFR) Pooam M, Nakayama M, Nishigaki C, Miyata H. Effect of 50-Hz sinusoidal magnetic field on the production of superoxide anion and the expression of heat-shock protein 70 in RAW264 cells. *Int J Chem* 9:23-36, 2017.

(E) (VT AE, IFR) Potenza L, Martinelli C, Polidori E, Zeppa S, Calcabrini C, Stocchi L, Sestili P, Stocchi V. Effects of a 300 mT static magnetic field on human umbilical vein endothelial cells. *Bioelectromagnetics.* 31(8):630-639, 2010.

(E) (VO, CE, IOD, IAO) Rageh MM, El-Gebaly RH, El-Bialy NS. Assessment of genotoxic and cytotoxic hazards in brain and bone marrow cells of newborn rats exposed to extremely low-frequency magnetic field. *J Biomed Biotechnol.* 2012:716023, 2012.

(E) (HU, AE, DOD) Raggi F, Vallesi G, Rufini S, Gizzi S, Ercolani E, Rossi R. ELF magnetic therapy and oxidative balance. *Electromagn Biol Med.* 27(4):325-339, 2008.

(E) (VO, AE, IAO) Rajabbeigi E, Ghanati F, Abdolmaleki P, Payez A. Antioxidant capacity of parsley cells (*Petroselinum crispum* L.) in relation to iron-induced ferritin levels and static magnetic field. *Electromagn Biol Med.* 32(4):430-441, 2013.

(E) (VO, CE, IX) Rauš Balind S, Selaković V, Radenović L, Prolić Z, Janać B. Extremely low frequency magnetic field (50 Hz, 0.5 mT) reduces oxidative stress in the brain of gerbils submitted to global cerebral ischemia. *PLoS One.* 9(2):e88921, 2014.

(E) (VT, AE, IAO) Reale M, De Lutiis MA, Patruno A, Speranza L, Felaco M, Grilli A, Macrì MA, Comani S, Conti P, Di Luzio S. Modulation of MCP-1 and iNOS by 50-Hz sinusoidal electromagnetic field. *Nitric Oxide.* 15(1):50-57, 2006.

- (E) (VT, AE, IFR) Reale M, Kamal MA, Patruno A, Costantini E, D'Angelo C, Pesce M, Greig NH. Neuronal cellular responses to extremely low frequency electromagnetic field exposure: implications regarding oxidative stress and neurodegeneration. *PLoS One*. 9(8):e104973, 2014.
- (E) (VO, CE, LI, IAO, DAO) Regoli F, Gorbi S, Machella N, Tedesco S, Benedetti M, Bocchetti R, Notti A, Fattorini D, Piva F, Principato G. Pro-oxidant effects of extremely low frequency electromagnetic fields in the land snail *Helix aspersa*. *Free Radic Biol Med*. 39(12):1620-1628, 2005.
- (E) (VT, AE, IFR) Rollwitz J, Lupke M, Simkó M. Fifty-hertz magnetic fields induce free radical formation in mouse bone marrow-derived promonocytes and macrophages. *Biochim Biophys Acta*. 1674(3):231-238, 2004.
- (NE) (IV, CE) Romeo S, Sannino A, Scarfi MR, Massa R, d'Angelo R, Zeni O. Lack of effects on key cellular parameters of MRC-5 human lung fibroblasts exposed to 370 mT static magnetic field. *Sci Rep*. 6:19398, 2016.
- (E) (VT, AE, IFR) Roy S, Noda Y, Eckert V, Traber MG, Mori A, Liburdy R, Packer L. The phorbol 12-myristate 13-acetate (PMA)-induced oxidative burst in rat peritoneal neutrophils is increased by a 0.1 mT (60 Hz) magnetic field. *FEBS Lett*. 376(3):164-166, 1995.
- (E) (VT, AE, IFR) Sadeghipour R, Ahmadian S, Bolouri B, Pazhang Y, Shafiezadeh M. Effects of extremely low-frequency pulsed electromagnetic fields on morphological and biochemical properties of human breast carcinoma cells (T47D). *Electromagn Biol Med*. 31(4):425-435, 2012.
- (E) (VT, CE, IOD, IAO, DAO) Sahebamei H, Abdolmaleki P, Ghanati F. Effects of magnetic field on the antioxidant enzyme activities of suspension-cultured tobacco cells. *Bioelectromagnetics*. 28(1):42-47, 2007.
- (E) (VO, CE, IFR) Salunke BP, Umathe SN, Chavan JG. Experimental evidence for involvement of nitric oxide in low frequency magnetic field induced obsessive compulsive disorder-like behavior. *Pharmacol Biochem Behav*. 122:273-278, 2014.

Santini SJ, Cordone V, Falone S, Mijit M, Tatone C, Amicarelli F, Di Emidio G. Role of mitochondria in the oxidative stress induced by electromagnetic fields: focus on reproductive systems. *Oxid Med Cell Longev*. 2018:5076271, 2018. (Review)

(E) (VO, CE, IOD, DAO) Seif F, Bayatiani MR, Ansarihadipour H, Habibi G, Sadelaji S. Protective properties of *Myrtus communis* extract against oxidative effects of extremely low-frequency magnetic fields on rat plasma and hemoglobin. *Int J Radiat Biol*. 2018 Nov 29:1-22. doi: 10.1080/09553002.2019.1542182. [Epub ahead of print]

(E) (VO, IOD, IAO, DAO) Seifirad S, Farzampour S, Nourbakhsh M, Amoli MM, Razzaghy-Azar M, Larijani B. Effects of extremely low frequency electromagnetic fields on paraoxonase serum activity and lipid peroxidation metabolites in rat. *J Diabetes Metab Disord*. 13(1):85, 2014.

(E) (VO, CE, IFR, IOD, IAO) Selaković V, Rauš Balind S, Radenović L, Prolić Z, Janać B. Age-dependent effects of ELF-MF on oxidative stress in the brain of mongolian gerbils. *Cell Biochem Biophys*. 66(3):513-521, 2013.

Seyhan N, Güler G. Review of in vivo static and ELF electric fields studies performed at Gazi Biophysics Department. *Electromagn Biol Med*. 25(4):307-323, 2006. (Review)

(E) (VO, CE, DAO) Sharifian A, Gharavi M, Pasalar P, Aminian O. Effect of extremely low frequency magnetic field on antioxidant activity in plasma and red blood cells in spot welders. *Int Arch Occup Environ Health*. 82(2):259-266, 2009.

(E) (VT, AE, IFR) Sherrard RM, Morellini N, Jourdan N, El-Esawi M, Arthaut LD, Niessner C, Rouyer F, Klarsfeld A, Doulazmi M, Witczak J, d'Harlingue A, Mariani J, Mclure I, Martino CF, Ahmad M. Low-intensity electromagnetic fields induce human cryptochrome to modulate intracellular reactive oxygen species. *PLoS Biol*. 16(10):e2006229, 2018.

(E) (VO, AE, IFR, IAO, DAO) Shine MB, Guruprasad KN, Anand A. Effect of stationary magnetic field strengths of 150 and 200 mT on reactive oxygen species production in soybean. *Bioelectromagnetics*. 33(5):428-437, 2012.

(E) (VO, CE, IFR, DFR, IAO, DAO) Shokrollahi S, Ghanati F, Sajedi RH, Sharifi M. Possible role of iron containing proteins in physiological responses of soybean to static magnetic field. *J Plant Physiol.* 226:163-171, 2018.

(E) (VT, AE, IFR) Simkó M, Droste S, Kriehuber R, Weiss DG. Stimulation of phagocytosis and free radical production in murine macrophages by 50 Hz electromagnetic fields. *Eur J Cell Biol.* 80(8):562-566, 2001.

Simkó M. Cell type specific redox status is responsible for diverse electromagnetic field effects. *Curr Med Chem.* 14(10):1141-1152, 2007. (Review)

Simkó M, Mattsson MO. Extremely low frequency electromagnetic fields as effectors of cellular responses in vitro: possible immune cell activation. *J Cell Biochem.* 93(1):83-92, 2004. (Review)

(E) (HU, AE, IAO, DFR) Sirmatel O, Sert C, Sirmatel F, Selek S, Yokus B. Total antioxidant capacity, total oxidant status and oxidative stress index in the men exposed to 1.5 T static magnetic field. *Gen Physiol Biophys.* 26(2):86-90, 2007a.

(E) (HU, AE, IFR) Sirmatel O, Sert C, Tümer C, Oztürk A, Bilgin M, Ziylan Z. Change of nitric oxide concentration in men exposed to a 1.5 T constant magnetic field. *Bioelectromagnetics.* 28(2):152-154, 2007b.

(E) (VT, AE, IFR) Solek P, Majchrowicz L, Bloniarz D, Krotoszynska E, Kozirowski M. Pulsed or continuous electromagnetic field induce p53/p21-mediated apoptotic signaling pathway in mouse spermatogenic cells in vitro and thus may affect male fertility. *Toxicology.* 382:84-92, 2017.

(E) (IV, AE, DFR) Song K, Im SH, Yoon YJ, Kim HM, Lee HJ, Park GS. A 60 Hz uniform electromagnetic field promotes human cell proliferation by decreasing intracellular reactive oxygen species levels. *PLoS One.* 13(7):e0199753, 2018.

(E) (VT, AE, IFR) Sullivan K, Balin AK, Allen RG. Effects of static magnetic fields on the growth of various types of human cells. *Bioelectromagnetics.* 32(2):140-147, 2011.

(E) (IV, AE, IFR, AO) Sun L, Chen L, Bai L, Xia Y, Yang X, Jiang W, Sun W. Reactive oxygen species mediates 50-Hz magnetic field-induced EGF receptor clustering via acid sphingomyelinase activation. *Int J Radiat Biol.* 94(7):678-684, 2018.

(E) (VT, AE, DFR) Sun YL, Chen ZH, Chen XH, Yin C, Li DJ, Ma XL, Zhao F, Zhang G, Shang P, Qian AR. Diamagnetic levitation promotes osteoclast differentiation from RAW264.7 cells. *IEEE Trans Biomed Eng.* 62(3):900-908, 2015.

(E) (VO, CE, IFR, IAO) Sun Y, Shi Z, Wang Y, Tang C, Liao Y, Yang C, Cai P. Coupling of oxidative stress responses to tricarboxylic acid cycle and prostaglandin E₂ alterations in *Caenorhabditis elegans* under extremely low-frequency electromagnetic field. *Int J Radiat Biol.* 2018 Oct 11:1-8. doi: 10.1080/09553002.2019.1524943. [Epub ahead of print]

(E) (VT, AE, IFR, AO) Tang R, Xu Y, Ma F, Ren J, Shen S, Du Y, Hou Y, Wang T. Extremely low frequency magnetic fields regulate differentiation of regulatory T cells: Potential role for ROS-mediated inhibition on AKT. *Bioelectromagnetics.* 37(2):89-98, 2016.

(E) (VO, CE, IX) Tasset I, Medina FJ, Jimena I, Agüera E, Gascón F, Feijóo M, Sánchez-López F, Luque E, Peña J, Drucker-Colín R, Túnez I. Neuroprotective effects of extremely low-frequency electromagnetic fields on a Huntington's disease rat model: effects on neurotrophic factors and neuronal density. *Neuroscience.* 209:54-63, 2012.

(E) (VO, CE, IOD, DAO) Tayefi H, Kiray A, Kiray M, Ergur BU, Bagriyanik HA, Pekcetin C, Fidan M, Ozogul C. The effects of prenatal and neonatal exposure to electromagnetic fields on infant rat myocardium. *Arch Med Sci.* 6(6):837-842, 2010.

(E) (VO, CE, IAO) Todorović D, Mirčić D, Ilijin L, Mrdaković M, Vlahović M, Prolić Z, Mataruga VP. Effect of magnetic fields on antioxidative defense and fitness-related traits of *Baculum extradentatum* (insecta, phasmatodea). *Bioelectromagnetics.* 33(3):265-273, 2012.

(E) (VO, CE, IAO) Todorović D, Ilijin L, Mrdaković M, Vlahović M, Filipović A, Grčić A, Perić-Mataruga V. Long-term exposure of cockroach *Blaptica dubia* (Insecta: Blaberidae) nymphs to magnetic fields of different

characteristics: Effects on antioxidant biomarkers and nymphal gut mass. *Int J Radiat Biol.* 95(8):1185-1193, 2019.

(NE) (VO, CE, IX) Túnez I, Drucker-Colín R, Jimena I, Medina FJ, Muñoz Mdel C, Peña J, Montilla P. Transcranial magnetic stimulation attenuates cell loss and oxidative damage in the striatum induced in the 3-nitropropionic model of Huntington's disease. *J Neurochem.* 97(3):619-630, 2006.

(NE) (VO, CE) Türközer Z, Güler G, Seyhan N. Effects of exposure to 50 Hz electric field at different strengths on oxidative stress and antioxidant enzyme activities in the brain tissue of guinea pigs. *Int J Radiat Biol.* 84(7):581-590, 2008.

(E) (VO, AE, TFR, DFR) Van Huizen AV, Morton JM, Kinsey LJ, Von Kannon DG, Saad MA, Birkholz TR, Czajka JM, Cyrus J, Barnes FS, Beane WS. Weak magnetic fields alter stem cell-mediated growth. *Sci Adv.* 5(1):eaau7201, 2019.

(NE) (VT, AE) Vannoni D, Albanese A, Battisti E, Aceto E, Giglioni S, Corallo C, Carta S, Ferrata P, Fioravanti A, Giordano N. In vitro exposure of human osteoarthritic chondrocytes to ELF fields and new therapeutic application of musically modulated electromagnetic fields: biological evidence. *J Biol Regul Homeost Agents.* 26(1):39-49, 2012.

(E) (VO, CE, DFR, DAO) Vignola MB, Dávila S, Cremonezzi D, Simes JC, Palma JA, Campana VR. Evaluation of inflammatory biomarkers associated with oxidative stress and histological assessment of magnetic therapy on experimental myopathy in rats. *Electromagn Biol Med.* 31(4):320-332. 2012.

(NE) (VT, AE, IAO) Villarini M, Gambelunghe A, Giustarini D, Ambrosini MV, Fatigoni C, Rossi R, Dominici L, Levorato S, Muzi G, Piobbico D, Mariucci G. No evidence of DNA damage by co-exposure to extremely low frequency magnetic fields and aluminum on neuroblastoma cell lines. *Mutat Res.* 823:11-21, 2017.

Vojtíšek M, Knotková J, Kasparová L, Svandová E, Markvartová V, Tůma J, Vozeh F, Patková J. Metal, EMF, and brain energy metabolism. *Electromagn Biol Med.* 28(2):188-193, 2009. (Review)

(E) (IV, AE, IFR, DFR) Wang D, Zhang L, Shao G, Yang S, Tao S, Fang K, Zhang X. 6-mT 0-120-Hz magnetic fields differentially affect cellular ATP levels. *Environ Sci Pollut Res Int.* 25(28):28237-28247, 2018.

Wang H, Zhang X. Magnetic fields and reactive oxygen species. *Int J Mol Sci.* 18(10), 2017, pii: E2175. doi: 10.3390/ijms18102175 Review. (Review)

Wang S, Luo J, Zhang Z, Dong D, Shen Y, Fang Y, Hu L, Liu M, Dai C, Peng S, Fang Z, Shang P. Iron and magnetic: new research direction of the ferroptosis-based cancer therapy. *Am J Cancer Res.* 8(10):1933-1946, 2018. (Review)

(E) (VT, AE, AO, IAO, AO) Wartenberg M, Wirtz N, Grob A, Niedermeier W, Hescheler J, Peters SC, Sauer H. Direct current electrical fields induce apoptosis in oral mucosa cancer cells by NADPH oxidase-derived reactive oxygen species. *Bioelectromagnetics.* 29(1):47-54, 2008.

(E) (VT, AE, IOD, IFR, AO) Wolf FI, Torsello A, Tedesco B, Fasanella S, Boninsegna A, D'Ascenzo M, Grassi C, Azzena GB, Cittadini A. 50-Hz extremely low frequency electromagnetic fields enhance cell proliferation and DNA damage: possible involvement of a redox mechanism. *Biochim Biophys Acta.* 1743(1-2):120-129, 2005.

(E) (VO, CE, IAO) Wu SX, Xu YQ, Di GQ, Jiang JH, Xin L, Wu TY. Influence of environmental static electric field on antioxidant enzymes activities in hepatocytes of mice. *Genet Mol Res.* 15(3), 2016. doi: 10.4238/gmr.15038800.

(E) (VT, AE, IFR, AO) Yang ML, Ye ZM. [Extremely low frequency electromagnetic field induces apoptosis of osteosarcoma cells via oxidative stress]. *Zhejiang Da Xue Xue Bao Yi Xue Ban.* 44(3):323-328, 2015. [Article in Chinese]

(E)(VO, IV, IFR) Yang LL, Zhou Y, Tian WD, Li HJ, Kang-Chu-Li, Miao X, An GZ, Wang XW, Guo GZ, Ding GR.: Electromagnetic pulse activated brain microglia via the p38 MAPK pathway. *Neurotoxicology.* 52:144-149, 2016.

(E) (VT, AE, IFR, AO) Yin C, Luo X, Duan Y, Duan W, Zhang H, He Y, Sun G, Sun X. Neuroprotective effects of lotus seedpod procyanidins on extremely low frequency electromagnetic field-induced neurotoxicity in primary cultured hippocampal neurons. *Biomed Pharmacother.* 82:628-639, 2016.

- (E) (VO, CE, IOD) Yokus B, Cakir DU, Akdag MZ, Sert C, Mete N. Oxidative DNA damage in rats exposed to extremely low frequency electromagnetic fields. *Free Radic Res.* 39(3):317-323, 2005.
- (E) (VO, CE, IOD) Yokus B, Akdag MZ, Dasdag S, Cakir DU, Kizil M. Extremely low frequency magnetic fields cause oxidative DNA damage in rats. *Int J Radiat Biol.* 84(10):789-795, 2008.
- (NE) (VT, AE) Yoon HE, Lee JS, Myung SH, Lee YS. Increased γ -H2AX by exposure to a 60-Hz magnetic fields combined with ionizing radiation, but not hydrogen peroxide, in non-tumorigenic human cell lines. *Int J Radiat Biol.* 90(4):291-298, 2014.
- (NE) (VO, AE, IX) Yoshikawa T, Tanigawa M, Tanigawa T, Imai A, Hongo H, Kondo M. Enhancement of nitric oxide generation by low frequency electromagnetic field. *Pathophysiology.* 7(2):131-135, 2000.
- (E) (VO, AE, DAO, IOD) Zeng L, Ji X, Zhang Y, Miao X, Zou C, Lang H, Zhang J, Li Y, Wang X, Qi H, Ren D, Guo G. MnSOD expression inhibited by electromagnetic pulse radiation in the rat testis. *Electromagn Biol Med.* 30(4):205-218, 2011
- (E) (IV, AE, CE, IFR) Zeng Y, Shen Y, Hong L, Chen Y, Shi X, Zeng Q, Yu P. Effects of single and repeated exposure to a 50-Hz 2-mT electromagnetic field on primary cultured hippocampal neurons. *Neurosci Bull.* 33(3):299-306, 2017.
- (E) (HU, CE, IOD, AO) Zhang D, Zhang Y, Zhu B, Zhang H, Sun Y, Sun C. Resveratrol may reverse the effects of long-term occupational exposure to electromagnetic fields on workers of a power plant. *Oncotarget.* 8(29):47497-47506, 2017.
- Zhang J, Ding C, Ren L, Zhou Y, Shang P. The effects of static magnetic fields on bone. *Prog Biophys Mol Biol.* 114(3):146-152, 2014. (Review)
- (E) (VT, AE, IFR, DRF) Zhang J, Ding C, Meng X, Shang P. Nitric oxide modulates the responses of osteoclast formation to static magnetic fields. *Electromagn Biol Med.* 37(1):23-34, 2018.

(E) (VO, AE, IX) Zhang ZY , Zhang J , Yang CJ, Lian HY, Yu H, Huang XM, Cai P. Coupling Mechanism of Electromagnetic Field and Thermal Stress on *Drosophila melanogaster*. PLoS One. 11(9):e0162675, 2016.

(E) (VT, AE, IFR) Zhao G, Chen S, Wang L, Zhao Y, Wang J, Wang X, Zhang W, Wu R, Wu L, Wu Y, Xu A. Cellular ATP content was decreased by a homogeneous 8.5 T static magnetic field exposure: role of reactive oxygen species. Bioelectromagnetics. 32(2):94-101, 2011.

(E) (VT, AE, IX, LI) Zmyslony M, Rajkowska E, Mamrot P, Politanski P, Jajte J. The effect of weak 50 Hz magnetic fields on the number of free oxygen radicals in rat lymphocytes in vitro. Bioelectromagnetics. 25(8):607-612, 2004a.

(E) (VT, AE, IX, LI) Zmyslony M, Palus J, Dziubałowska E, Politański P, Mamrot P, Rajkowska E, Kameduła M. Effects of in vitro exposure to power frequency magnetic fields on UV-induced DNA damage of rat lymphocytes. Bioelectromagnetics. 25(7):560-562, 2004b.

(E) (VT, AE, IOD, IAO) Zwirska-Korczala K, Adamczyk-Sowa M, Polaniak R, Sowa P, Birkner E, Drzazga Z, Brzozowski T, Konturek SJ. Influence of extremely-low-frequency magnetic field on antioxidative melatonin properties in AT478 murine squamous cell carcinoma culture. Biol Trace Elem Res. 102(1-3):227-243, 2004.
